## Module 3: Comparative Sequence Analysis and Identification of Regulatory Elements

## Aims

- Overview of comparative sequence analysis and sequences available from different organisms for homologous gene identification
- Identify putative paralogous and orthologous genes in Ensembl
- View phylogenetic trees
- Compare genome sequences from different organisms in UCSC
- Identify evolutionary conserved sequences (ECRs) in ECR browser and VISTA Enhancer Browser

## **Comparative Sequence Analysis**

Comparative sequence analysis is a powerful method for aiding human gene identification, inferring function of a gene's product, and identifying novel functional elements such as those involved in transcriptional regulation. This is because biologically important regions of the genome are, generally, under selective constraint. Comparing the genome sequences from a variety of organisms may facilitate the identification of functionally significant units in the human genome.

The information that can be inferred when comparing sequences is dependent on the evolutionary distance between the two organisms. Organisms that are closely related are more likely to share a higher degree of sequence similarity. Organisms more distantly related to human, such as yeast and worm, share less sequence similarity and are likely to show sequence conservation only in coding regions. This may also be true for distantly related vertebrates such as fish. More closely related organisms, such as mouse, are likely to be conserved in coding regions, and other functional elements such as regulatory sequences. However, the closer the evolutionary relationship with human, the more 'sequence noise' is likely to arise where non-functional sequence appears similar because insufficient time has elapsed for the two sequences to diverge.

## **Evolutionarily Related Gene Sequences**

Homologous genes are derived from a common ancestor and may retain a similar sequence or function. In general, homologous genes can be divided into two classes:

1) Orthologues are genes that often perform the same function in different organisms. They are defined as being homologous genes in different organisms derived from the same gene during speciation. In general, their sequence similarity reflects the amount of time since they diverged from a common ancestor (i.e. the less time that has elapsed since divergence, the greater the sequence similarity between the two genes).

**2) Paralogues** are gene families that are present within a single species. Often they arise by duplication. These genes are not under the same pressure to maintain their function so that one copy may acquire a novel function.

\*note: The terms orthologous and paralogous can apply to sequence without genes as well!



Figure 1 - Homologous Gene Sequences

## Identifying evolutionarily related gene sequences – where to start...

Most searches for orthologous genes begin with BLAST searches. The type of search that you should perform depends upon what information you have at your disposal. Protein sequence based queries generally find more distantly related matches because of the redundancy in the genetic code (i.e. some amino acids are encoded by more than one codon). Nucleotide searches using the discontiguous megablast parameters are also very useful. We recommend that you survey a number of different databases using a number of different search parameters to obtain the most informative results.

## **BE CAUTIOUS: uncertain Orthologues**

You may encounter gene sequences that appear to be orthologous and may be derived from the same ancestor but no longer perform the same biological function (for example genes 1 and 3 in Figure 1). If you choose to analyze such sequences, the sensitivity and specificity of your search will be reduced and it may not yield any informative results.

For example, the gene for bone morphogenetic protein 8 (*BMP8*) was duplicated in a common ancestor of human and mouse, giving rise to *BMP8a* and *BMP8b* (see Figure below). BLAST analysis of these four sequences yields quite confusing results. Human and mouse *BMP8a* are reciprocal best alignments using both nucleotide and protein sequences to search. In contrast, both the nucleotide and protein sequences of mouse *Bmp8b* align best to their human *BMP8a* counterparts. Human *BMP8b* mRNA aligns best to mouse *Bmp8b* mRNA, but human BMP8b protein aligns best to mouse Bmp8a protein, while mouse protein *Bmp8b* aligns best to human protein *BMP8a* (Nardone*et al.,* 2004).

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Figure 2 - Uncertain BMP8a and BMP8b orthologues.

Therefore, we recommend that you perform the following steps to confirm the true functionality and relatedness of your gene sequences. Much of this information can be obtained from genome browsers such as Ensembl, NCBI or the UCSC.

1) Identify any other paralogues that may affect your analysis. This can be achieved by performing a BLAST search using your sequence against its source genome, or using the self-chain track at the UCSC genome browser.

2) Confirm the percent identity (similarity) at both the nucleotide and protein level between paralogous and orthologous sequences to ensure that you are analyzing the most closely related sequences.

3) Perform evolutionary analyses of nucleotide/protein sequence (phylogeny). In contrast to similarity-based methods such as BLAST, phylogenetic methods can better take into account the effects of repeated substitutions at one site and variable rates of evolution among sequences. Multiple genes are placed in an evolutionary tree representing genealogical relationships.

4) Compare the exon/intron structure of your orthologous genes.Evolutionary related genes often share a similar gene structure.

5) Examine the chromosomal context of the two orthologous genes. Closely related species, such as human and mouse often have large

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conserved segments and therefore neighbouring genes are also shared between the two species.

## **Comparative Genome Analysis**

Comparing the DNA sequences of different species is a powerful method for decoding genomic information. This is because functional sequences tend to evolve at a slower rate than non-functional sequences (see Figure 3).



**Figure 3 - Diverging sequences. Miller** *et al***, 2004. Annu Rev Genomics Hum Genet.** Immediately after speciation the sequence is 100% identical (see graph on left).Over time, regions under little or no selective pressure, such as introns, are saturated with mutations, whereas regions under negative selection, such as most exons, retain a higherpercent identity (see graph on right).

By comparing the genomic sequences of several species at different evolutionary distances it is possible to identify coding sequences, conserved non-coding sequences, and those sequences that are unique to humans or other species.

When two species diverge from a common ancestor, those sequences that maintain their original function are likely to remain conserved in both species throughout their subsequent independent evolution. Therefore, comparing sequences in different species is a powerful tool for increasing the confidence of a predicted functional unit, or identifying novel functional units (e.g. human, mouse and zebrafish).

The sequence of many genomes has been generated using a combination of the clone-by-clone method (adopted for generating the human genome sequence by the public effort) and whole genome shotgun (WGS - used by Celera to generate the

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sequence of the fruit fly and their version of the human genome sequence). Finished and unfinished genome assemblies are currently available for many vertebrates.



The key genomes being used to predict conserved elements are shown from the UCSC website.

# Figure 4 - The genomes that have been aligned to predict conserved elements at the UCSC genome browser.

In addition to many completely sequenced genomes, low coverage sequence for numerous genomes is currently available and more are in queue to be processed. Many new sequencing platforms offered by Illumina, Roche (454) and Agencourt/ABI (SOLiD) are offering faster ways to obtain many new partial and whole genome sequences for much less cost. The draft sequence of the giant panda genome was the first to use only next generation sequencing (Li *et al* Nature 2010).

In general, greater evolutionary distance between the species is reflected by more divergent sequences and fewer shared functional units (see table below). Comparing sequences that diverged from a common ancestor approximately 450 million years ago (mya) (e.g. human and fish), aids the identification of coding sequences. Conserved non-coding regions are generally not identified. If the evolutionary distance between the two species is reduced to approximately 90 mya (e.g. human and mouse), both non-coding and coding units are commonly conserved. However there can be background noise so comparing a number of different mammalian species (including the more distantly related marsupials) is useful. A large number of features are conserved between recently evolved species such as human and chimpanzee. The inclusion of a closely related species in a comparative analysis makes it possible to identify those genomic sequences that may be responsible for traits that are unique to the reference species.

Human vs.	Chimpanzee	Mouse	Opossum	Fish
Size (Gbp)	3.0	2.5	3.6	0.4
Time since divergence	~5 MYA	~65MYA	~150 MYA	~450 MYA
Sequence conservation (in coding regions)	>99%	~80%	~75%	~65%
Aids identification of 	Recently changed sequences and genomic rearrangements	Both coding and non- coding sequences	Both coding and non- coding sequences	Primary coding sequences
Background noise	HIGH	MODERATE	LOWER	LOW

## Table 1 - Selection of Species for DNA Comparisons

## WORKED EXAMPLES:

## 1. Identifying orthologues and paralogues in Ensembl.

The easiest and most common method used to identify homologous sequences exploits the sequence conservation between related genes. This can be identified using sequence similarity searches (i.e. BLAST searches) against nucleotide or protein databases (see previous modules). However, in many cases, the genome browsers such as NCBI, Ensembl and the UCSC have already done the hard work for you. In this example, explore the data available in Ensembl.





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some browsers, a second Jalview button may appear. Click JalView for family members.

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ENSGALP00000030066/1-1398	ADT	RAMAKEROKKDNHN	L	IERRRRYNINYRI	KEL	GTLII	PKSND-PD	MRWNKGTILKASVE	
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Step 6: Return to the gene view page for *MITF* and view. This is a different way of getting to the same answer.

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## Orthologues

Orthologues are the same gene but in different species. To investigate the evolution of a gene we can investigate the relationships between orthologues and paralogues in all the species for which genomic sequence is available





Maximum likelihood phylogenetic tree drawn using **PHYML**. Red squares represent duplication nodes, while blue squares represent speciation nodes. When the branch length is too long for the display, they are shortened and displayed as dashed lines with color depending on the branch length. The green bars at the right of the tree are a schematic representation of the multiple alignment of the peptides made using **MUSCLE**. Full boxes indicate matches/mismatches, open boxes indicate gaps in the alignment.

Ensembl uses a pipeline to predict orthologues and paralogues in which "maximum likelihood phylogenetic gene trees (generated by TreeBeST) play a central role". However, **BE WARNED** as some of the orthologues predicted in the less characterised genomes are not always well annotated.



## 2. Comparing genome sequences in the UCSC.

We will next use the UCSC genome browser to identify evolutionarily conserved regions and then investigate them to see if they have any epigenetic signatures that suggest they have regulatory roles in the genome.











Zoom in on the ECR. Is it associated with any transcription factor binding sites? Switch on TFBS conserved Track to "pack" (Regulation section).



Yes, the ECR has a number of conserved transcription factor binding sites. This track shows TFBS that are conserved between human, mouse and rat.

## Adding your own track to UCSC.

It is really easy to make and upload data files for your own tracks to UCSC. We are going to add a very short bed file containing putative enhancers identified next to the H19 non-coding RNA gene. The file also contains the known control region for H19 and IGF2 genes. The file is called UCSC-enhancers\_track.bed.

```
browser position chrl1:200000-2030000
track name=PutativeEnhancers description="Putative_Enhancers" color=250,0,0
chrl1 2013859 2014040 CS1
chrl1 2012311 2012615 CS2
chrl1 2011146 2011294 CS3
chrl1 2009454 2009727 CS4
chrl1 2001080 2001174 CS6
chrl1 2021124 2023944 H19-DMR
```

This is a very basic track with only 5 regions. The format has a description line including track name, description and in this case colour. This also includes the position I am interested in in the genome. Below you need at least 3 columns chromosome, start and end. I have also included a name column. Details of all of the file types you can upload can be found on the Add Custom Tracks page



Add Custom Tracks
clade Mammal : genome Human : assembly Feb. 2009 (CRCh37/hg19) : Display your own data as custom annotation tracks in the browser. Data must be formatted in <u>BED</u> , <u>bigBed</u> , <u>bedGraph</u> , <u>GFF</u> , <u>GTF</u> , <u>WIG</u> , <u>bigWig</u> , <u>MAF</u> , <u>BAM</u> , <u>BED detail</u> , <u>Personal Genome SNP</u> , <u>VCF</u> , <u>broadPeak</u> , <u>narrowPeak</u> , or <u>PSL</u> formats. To configure the display, set <u>track</u> and <u>browser</u> line attributes as described in the <u>User's Guide</u> . Data in the bigBed, <u>bigWig</u> , <u>BAM</u> and VCF formats can be provided via only a URL or embedded in a track line in the box below. Publicly available custom tracks are listed <u>here</u> . Examples are <u>here</u> .
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Click here for an HTML document template that may be used for Genome Browser track descriptions.

This is a short file so it can just be pasted in.

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go to table brow	ser
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Make sure the display is set to "full" for your custom track, otherwise it won't display the feature names.





These regions, which are viewed here as red bars, can be explored to see if they lie in areas of the genome that act as regulatory elements.

It is possible to save your session in UCSC so you can come back to it at a later date. Go to the Sessions page, which is under my data. On your first visit you can request an account.

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#### Sign in to UCSC Genome Bioinformatics

#### Login

#### Create an account

Signing in enables you to save current settings into a named session, and then restore settings from the session later. If you wish, you can share named sessions with other users.

#### Session Management

See the Sessions User's Guide for more information about this tool.

Click here to reset the browser user interface settings to their defaults.

If you sign in, you will also have the option to save named sessions.

#### **Save Settings**

Save current settings to a local file:

file: \_\_\_\_\_\_\_ file type returned: plain text + \_\_\_\_\_\_ submit

(leave file blank to get output in browser window)

#### **Restore Settings**

Use settings from an	other user's saved session:		
user:	session name:	sub	mit
Use settings from a	ocal file:	Browse	submit
Use settings from a	URL (http://, ftp://):	submit	

You can then save and share your sessions.

## 3. Comparing genome sequences in the ECR browser and identifying potential transcription factor binding sites.

The Comparative Genomics Developments website is a very useful resource comparing genomes to decipher the code of gene regulation. It encompasses much of the historical databases for genome alignments such as VISTA, rVISTA and zPicture. In this exercise we will explore the WWOX gene for conserved elements that may have regulatory function. But remember, conserved elements could be anything.



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chr16:78133327-79246564	WWOX	WW domain-containing oxidoreductase isoform 2	
UCSC genes		·····	
chr16:78133327-79246564	WWOX	Homo sapiens WW domain containing oxidoreductase (WWC mRNA.	DX), transcript variant 1,
chr16:78133327-78312593	WWOX	Homo sapiens WW domain containing oxidoreductase (WWC mRNA.	DX), transcript variant 2,
chr16:78133327-78134096	WWOX	Homo sapiens WW domain containing oxidoreductase (WWC mRNA.	DX), transcript variant 3,
chr16:78133327-79246564	WWOX	Homo sapiens WW domain containing oxidoreductase (WWC mRNA.	DX), transcript variant 1,
chr16:78133327-79246564	WWOX	Homo sapiens WW domain containing oxidoreductase (WWC mRNA.	DX), transcript variant 2,
chr16:78133327-79246564	WWOX	Homo sapiens WW domain containing oxidoreductase (WWC mRNA.	DX), transcript variant 1,
MGC genes			
chr16:78133597-79246564	BC003184	WWOX	



Step 4: The database is based on original VISTA plots and contains gene information imported from the UCSC. Be careful because it may not be reflecting the same genome build. On the right hand side, you can remove or add organisms (add cow and zebrafish). Click instructions at the top to get the helpful key shown below. Change the parameters using the [change] function.



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ECR B Para [ch	Browser on Human (hg19) <u>http://ecrbrow</u> ECR ECR Layer ameters: Graph length similarity height hange smooth 100 70 55	Coordinate system relative	ne or position (chrN.from-to) Ir16:78133327-79246564 <b>Submit</b> NOME ALIGNMENT: Align your sequence to a genome	
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Step 6: There is one very highly conserved ECR in intron 8 (the large final intron). It's easier to see if you zoom out first! Zoom in a bit and you may have to re-centre. Then mouse over the ECR on the fugu track. This will bring up an alignment between the two species.

If you find it tricky to get to this ECR, simply open ECR browser from the following co-ordinates: chr16:78464965-78550000

**RVISTA** 



DOWNLOADING



Step 8. You are now in rVISTA, but importantly, all the parameters have been filled in for you, and all you need to do is select the TFBS of interest. RVISTA TRANSCRIPTION FACTORS

				Total r	umber of tran	scription factor	families: 467
		SELE	CT TRANSCR	IPTION FAC	TORS		
SELECT SEPAR	ATE TRANSCRI	PTION FACTO	RS				
Α							
ACAAT_B	AFP1_Q6	AHR	AHRARNT	AHRHIF_Q6	AHR_Q5	AIRE	ALPHACP1
ALX4	AMEF2_Q6	AML1	AML1_Q6	AML_Q6	AP1	AP1FJ_Q2	AP1_C
■ <u>AP1_Q2</u>	<u>AP1_Q4</u>	■ <u>AP1_Q6</u>	AP2ALPHA	AP2GAMMA	AP2REP	AP2 Q3	■ <u>AP2_Q6</u>
■ <u>AP3_Q6</u>	<u>AP4</u>	AP4_Q5	<u>AP4_Q6</u>	APOLYA B	⊟ <u>AR</u>	AREB6	ARNT
ARP1	AR_Q2	■ <u>AR_Q6</u>	ATATA_B	ATF	ATF1_Q6	ATF3_Q6	ATF4_Q2
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BACH1	BACH2	BARBIE	BEL1_B	BLIMP1_Q6	BRACH	BRCA	BRN2
CAAT	CAAT_C	CACBINDING		CACD	□ <u>CAP</u>	CART1	E CBF
CDC5		CDPCR1	CDPCR3	CDPCR3HD	CDX2_Q5	CDXA	CDX Q5
CEBP	CEBPA	CEBPB	CEBPDELTA	CEBPGAMMA	CEBP_C	CEBP_Q2	CEBP_Q3
CETS168_Q6	CETS1P54	📃 <u>СНСН</u>	CHOP	CHX10	□ <u>CIZ</u>	CLOCKBMAL	
CMAF	CMYB	COMP1			COUPTF_Q6	COUP_DR1_Q	CP2
CREB	CREBATF_Q6	CREBP1	CREBP1CJUN	CREBP1_Q2	CREB_Q2	CREB Q3	CREB_Q4
CREL	CRX_Q4						
D E							



Request ID: <u>xbr04142010123759740</u>		
Summary: <u>54 conserved</u> and <u>54 aligned</u> transcription fa	ctor binding sites (TFBS) were identified	
Dynamic visualization: Dynamically overlaw TFBS prediction with the and perform clustering	conservation profile	
Alignment: Highlight TFBS positions in the alignment	40 50 \ATAAGAGATAATAATCTATT ::              :: 3CT GAGATAATAATCTAAG 3 60	Step 10: Choose th dynamic visualization
Binding sites in the input sequences: <u>96 TFBS</u> detected in the base sequence <u>89 TFBS</u> detected in the second sequence		

RVISTA VISUALIZATION & CLUSTERING





Can we find any other functional information about this ECR?





## Human element [ hs12 ]

Position: chr16:78,510,608-78,511,944 (UCSC browser) Source: Lawrence Berkeley National Laboratory

Flanking genes: WWOX - MAF

#### **Expression Pattern**

forebrain (9 out of 11 embryos)

hindbrain (rhombencephalon) (9 out of 11 embryos)

Embryo 1



Embryo 3









## Tasks:

- 1. Ensembl:
  - a. Identify the *Bax* gene in Zebrafish hint: you may not be able find it by searching in the zebrafish Ensembl.
  - b. Can you identify any predicted paralogues
  - c. View the orthologues for the zebrafish Bax gene
  - d. View the Gene tree view for Bax
- 2. UCSC genome browser:
  - a. Find the ARID1A gene in the genome.
  - b. Are there any non-coding ECRs within or close to this gene?

c. A recent paper has identified putative heart enhancers in developing mouse heart via p300 ChIP (Blow *et al.* Nat Genet. 2010 September; 42(9): 806–810). On your computer is a file of the coordinates of these putative elements (p300\_Heart). Read this file in to UCSC and see if any of the peaks coincide with the regions identified above.

- 3. ECR browser. Using the Human ZAK gene:
  - a. View the gene in the ECR browser
  - b. Can you find non-coding ECRs?
  - c. Set up rVISTA analysis between human and mouse on the most conserved ECR. Also try looking at the conserved TFBS for this ECR at UCSC
  - d. Is there any evidence for this being a regulatory element in the Vista Enhancer browser?

## Answers:

1. Ensembl:

a. Identify the *Bax* gene in Zebrafish – hint: you may not be able find it by searching in the zebrafish Ensembl.

Search for the human *BAX* gene and view the orthologues. There will be a zebrafish orthologue called *baxa*. Interestingly there are two copies of *BAX* in the zebrafish genome - it appears to have been tandemly duplicated *baxa* and *baxb*.

Show/hide columns					Filter	
Туре	Ancestral taxonomy	Ensembl identifier & gene name	Compare	Location	Target %id	Query %id
Paralogue (within species)	Danio rerio	ENSDARG0000089129 baxa bci2-associated X protein, a [Source-ZFIN;Acc:ZDB-GENE-000511-6]	<ul> <li>Region Comparison</li> <li>Alignment (protein)</li> <li>Alignment (cDNA)</li> </ul>	<u>3:37761255-37768582:-1</u>	50	52
Paralogue (within species)	Euteleostomi	ENSDARG00000030881 baxb bcl2-associated X protein, b [Source:2FIN:Acc:2DB- GENE-050227-21]	<ul> <li>Region Comparison</li> <li>Alignment (protein)</li> <li>Alignment (cDNA)</li> </ul>	<u>3:32211782-32216892:-1</u>	20	22
Paralogue (within species)	Euteleostomi	ENSDARG0000089995 BX511080.2 Uncharacterized protein [Source: UniProtKB/TrEMBL (E7F560)]	<ul> <li>Region Comparison</li> <li>Alignment (protein)</li> <li>Alignment (cDNA)</li> </ul>	<u>3:32204378-32208411:-1</u>	19	16
Paralogue (within species)	Euteleostomi	ENSDARG0000068102 zgc:153993 zgc:153993 [Source:ZFIN;Acc:ZDB- GENE-060029-176]	<ul> <li>Region Comparison</li> <li>Alignment (protein)</li> <li>Alignment (cDNA)</li> </ul>	<u>7:54755948-54779600:-1</u>	32	34

b. Can you identify any predicted paralogues for baxa?

c. View the orthologues for the zebrafish Bax gene

There are 55 orthologues

## d. View the Gene tree image for Bax



You can expand the image to view the full tree, and alter the display parameters under configure this page.

## 2. UCSC genome browser

a. Find the ARID1A gene in the human genome.

## ARID1A is found at chr1:27022522-27108601

## b. Are there any non-coding ECRs within or close to this gene?



Yes there are a number of non exonic ECRs

c. A recent paper has identified putative heart enhancers in developing mouse heart via p300 ChIP (Blow et al Nat Genet. 2010 September; 42(9): 806–810). On your computer is a file of the coordinates of these putative elements (p300\_Heart). Read this file in to UCSC and see if any of the peaks coincide with the regions identified above.



The file contained 3 peaks all of which correspond to heart H3K27ac peaks in heart derived tissues. Together these data suggest that these regions act as enhancers in the mammalian heart.

## 3. ECR browser. Using either the Human ZAK gene:



## a. View the gene in the ECR browser

b. Can you find non-coding ECRs?

There are a couple of non-coding ECRs that may be real. There is a really well conserved ECR in intron 14 of *ZAK*.

c. Set up rVISTA analysis between human and mouse

	ZAK	
MEF2	······································	CONSERVED
HI_Q2		
2F1_Q4		
YF1	1	
SRF_Q4 22F_Q4		
2F_Q6	i la seconda de la se Seconda de la seconda de la	
JATAI		
JATA2 JATA3		
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R_Q6		
COUPTF_	26	
F3R PFARG		
T3R_Q6	HA 06	
INF4_Q6		
REBPICJ	1_Q6 UN	
INF4_DR	L_Q3 02	
DRI_Q3		
SOX9_B1		
SOX5 DCT1	in a second	
OCT_C	II. II.	
OCT_Q6	įi	
DX_Q5		
EBPB		
EBPA		
DP	1	
MYB CARTI		
OCTL Q6		
DCT4	1 J I	
ICFIIMAI MAF_Q6	7G	
TA_Q2	P = 1	
ITTF1_Q3		
DELTAEF		
IMEF2_Q MEF2_Q6	6	
BACHI		
VFE2		
AP1FJ_Q2 AP1_Q2		
4P1_Q4 4P1_Q6		
BACH2		
TAXCREB		
RF_Q6	111	
BLIMP1_Q ÆLIOSA	26	
ENI		
UNI		
ALX4 PITI_Q6		
PBX1 PAX4		
APOLYA_	B C C C C C C C C C C C C C C C C C C C	
CSBP_Q6		
E4BP4 EVI1		
OTX_Q1 BRN2		
DXA	ī,	
MSX1	<u>1</u>	
MYCMAN MYOD		
BOX_Q6		
JSF_Q6		
JSF_C		
AML1_Q6		
REBI	AR	
R_Q2		
R_Q6	t t	
CETS1P54 STAT5A		
TAT6	06	
38		
VKX62_Q	2	
IOXA4_Q BRACH	2	
ATF4_Q2 ATF3_O6		
MAF		
0X04	11 I.	
MEF2_Q	<b>6</b> 26	
RY		

244 conserved transcription factor binding sites were identified between human and mouse.

This is quite meaningless; it is more informative to see which TFBS are conserved between more than 2 species. You can also look at the region in UCSC and see which conserved transcription factor binding sites were identified between human, mouse and rat



d. Is there any evidence for this being a regulatory element in the Vista Enhancer browser?

Yes - there is evidence for enhancer activity in the forebrain, midbrain and neural tube.

