## **Hybrid Capture protocol 2b**

# Solution hybridisation (Agilent SureSelect protocol)

This protocol is for hybridization of adapter-ligated or PCR-amplified library DNA, so must be performed after Hybrid Capture Protocol 1.

# Sample preparation

Following SPRI bead cleanup (see Hybrid Capture Protocol 1), lyophilize 500 ng library + 7.5  $\mu$ g C<sub>0</sub>t1 DNA using a SpeedVac..

- 1. Add 3.4 μl of molecular biology grade water to each sample to rehydrate.
- 2. Vortex and spin down.
- 3. To each sample, add:

```
SureSelect Block #1 2.5 µl
SureSelect Block #2 2.5 µl
SureSelect Block #3 0.6 µl
```

4. Prepare Hybridization Buffer as follows. Volume for 1 capture:

```
SureSelect Hyb #1 25 \mul
SureSelect Hyb #2 1 \mul
SureSelect Hyb #3 10 \mul
SureSelect Hyb #4 13 \mul
```

Note: Do NOT keep on ice.

- 5. Incubate 40  $\mu$ l of Hybridization Buffer and library (from step 3.) at 95 °C for 5 minutes and 65 °C for at least 5 minutes. Keep at 65 °C until RNA baits are prepared (see below).
- 6. Dilute RNAse Block in 1:1 ratio with nuclease-free water and add 1  $\mu$ l to 5  $\mu$ l (500 ng) of RNA baits. Incubate for 2 min at 65 °C.

#### Hybridization

- 1. Mix 13 μl of hybridization buffer with RNA baits.
- 2. Add the 9 μl DNA library to the hyb buffer RNA bait mixture.

3. Seal plate with the Greiner film and incubate for 24 hours at 65 °C with a heated lid at 105 °C.

## Washing and elution

#### Selection with magnetic beads

- 1. Prepare magnetic beads by washing with SureSelect Binding buffer 3 times and resuspend in 200  $\mu$ l of SureSelect Binding buffer. Incubate the hybrid-capture-bead solution on a Nutator for 30 minutes at room temperature (20 °C).
- 2. Mix hyb. mixture with beads and separate the beads by removing the supernatant.
- 3. Resuspend beads in SureSelect Wash Buffer #1.
- 4. Incubate the samples for 15 minutes at room temperature. Separate the beads and buffer on a Dynal magnetic separator and remove the supernatant.
- 5. Mix the beads in pre-warmed (65 °C) 500  $\mu$ L SureSelect Wash Buffer #2. Incubate the samples for 10 minutes at 65 °C. Remove wash buffer and repeat these steps for 3 times.
- 6. Mix the beads in 50  $\mu$ L SureSelect Elution Buffer. Incubate the samples for 10 minutes at room temperature. Separate the beads and buffer on a Dynal magnetic separator. Add 50  $\mu$ L of SureSelect Neutralization Buffer.
- 7. Desalt the capture solution with a Qiagen MinElute PCR purification column, eluting in  $34~\mu L$  buffer EB.

Proceed to the Post-hyb PCR (Hybrid Capture Protocol 3).