

Gene: Slc29a3

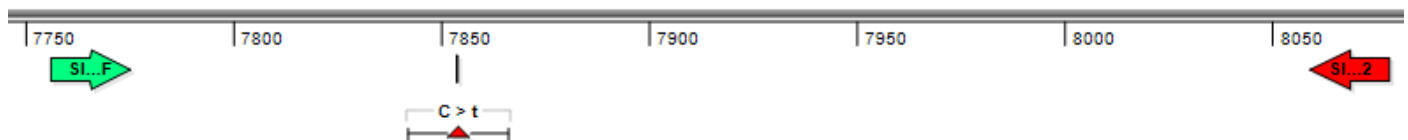
Colony prefix: DARW

Allele: *Slc29a3*^{em1(IMPC)Wtsi}

Allele type: Crispr/Cas9 mediated Point mutation

Allele information: E447K

Further information about the allele can be found on the 'International Mouse Phenotyping Consortium' (IMPC) web site at <http://www.mousephenotype.org/>



Mouse QC information

| | | | |
|------------------------------|----|------------------------------|------|
| Loss of WT Allele (LoA) qPCR | na | Mutation Sequence confirmed | pass |
| Mutant Specific SR-PCR | na | Off-target analysis complete | na |

Guide RNAs and mutant oligos used in initial experiment

| Sequence | Chr | Chr Start | Chr End |
|--|-----|-----------|----------|
| GTGGCCTCAGCCAGCTCCCGGG | 10 | 60715918 | 60715940 |
| GGTACCTCAGCACGCTGGTGCTCATCTATGGGCCCAAGATTG TGCCCCGGGAGCTGGCTAAGGCCACCAAGTGTGTGATGTTGTT CTATATGTCTGTGGGCTTGATGCTGGGCTCAGCCTGC | 10 | 60715863 | 60715985 |

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Mutant allele sequence:

CGCCCCCTCCTAGATAAAGTGTTCAAGCAGGGCCGCGCAGGCTGAGCCCAGCATCAAGCCCACAGACATATAGAACAACATCAC
AAACTGGTGGCCT[C/t]AGCCAGCTCCCGGGGCACAATCTGGGCCATAGATGAGCACCAGCGTGCTGAGGTAGCCGTTGCT
GAGCCCCAGGAGGCAGGTGAAAAGCACTGGGTAGATGTCCGACTGGAAAAGCACCTTGGTCAAGTGTGAGCGCGGCTGGTAG
TTACAGAGCAAGAAGAGAGGCCACAAGGCAGAACCGAGAGACCACCAGTCCGGGGAGCAGCTTGCTCCTAGGACCTG

Genotyping by end-point PCR

PCRs primer pairs and expected size bands

| Assay Type | Assay | Forward Primer | Reverse Primer | Expected Size Band (bp) |
|--------------|------------|-----------------|------------------|-------------------------|
| Standard PCR | Screening* | Slc29a3_PM_WT_F | Slc29a3_PM_WT_R2 | 323 |

*The screening PCR flanks the SNP region and can be used for sequence verification of the allele. The PCR will not distinguish wild type from mutant mice, however, as a product will be amplified in all cases.

We recommend that mice are sequence-verified with the screening primers to confirm the genotyping qPCR results when establishing the colony, in case of any cross-talk between the assays.

Primer sequences

| Primer Name | Primer Sequence (5' > 3') |
|------------------|---------------------------|
| Slc29a3_PM_WT_F | CGCCCCCTCCTAGATAAAGT |
| Slc29a3_PM_WT_R2 | CAGGTCCTAGGAGCAAGCTG |

Reaction setup

| Reagent | µl |
|---------------------------|------|
| DNA (~50-100 ng) | 1 |
| 10x Buffer | 2 |
| MgCl ₂ (50 mM) | 0.6 |
| Platinum Taq (Invitrogen) | 0.2 |
| dNTPs (100 mM) | 0.2 |
| Primer 1 (10 µM) | 0.4 |
| Primer 2 (10 µM) | 0.4 |
| ddH ₂ O | 15.2 |
| Total | 20 |

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Amplification conditions

| Step | Conditions | Time |
|------|-----------------------|----------|
| 1 | 94°C | 5 min |
| 2 | 94°C | 30 sec |
| 3 | 58°C | 30 sec |
| 4 | 72°C | 1:30 sec |
| 5 | Go to '2' + 34 cycles | - |
| 6 | 72°C | 5 min |
| 7 | 12°C | Forever |

Genotyping by SNP qPCR

Primers for LoA qPCR assay

| Gene | Source | Forward Primer Seq. | Reverse Primer Seq. | Probe Primer Seq. |
|---------|-------------------|-------------------------------|---------------------|---|
| Slc29a3 | Life Technologies | GCATCAAGCCCACAGACA TATAGAA | GTCATCTATGGGCCAAGA | [VIC]TGGCTGAGGCCACC [FAM]CTGGCTAAGGCCACC |

Reactions are performed in a 10µl volume using an Applied Biosystems 7900HT Fast Real-Time PCR System or Applied Biosystems Vii7 with DNA prepared using the Sample-to-SNP™ kit (Applied Biosystems) from mouse ear biopsies. GTXpress™ buffer is also used (Applied Biosystems).

| Reagent | µl |
|---------------------|------|
| 2x GTXpress™ buffer | 5 |
| 40x target assay | 0.25 |
| ddH2O | 3.75 |
| DNA | 1 |

Amplification conditions

| Step | Conditions | Time |
|-----------|----------------|--------|
| Pre-read | 60°C | 30 sec |
| 1 | 95°C | 20 sec |
| 2 | 95°C | 10 sec |
| 3 | 60°C | 30 sec |
| 4 | Go to '2' + 34 | - |
| Post-read | 60°C | 30 sec |

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Links to information and frequently asked questions

MGP mouse phenotype data:

<http://www.mousephenotype.org>

How the "critical" exon is decided:

<http://www.i-dcc.org/kb/entry/102/>

Relevant publications

White, J.K., Gerdin, A.-K., Karp, N.A., Ryder, E., Buljan, M., Bussell, J.N., Salisbury, J., Clare, S., Ingham, N.J., Podrini, C., et al. (2013). Genome-wide Generation and Systematic Phenotyping of Knockout Mice Reveals New Roles for Many Genes. *Cell* 154, 452–464.

Mali P, Yang L, Esvelt KM, et al (2013) RNA-guided human genome engineering via Cas9. *Science* 339:823–6. doi: 10.1126/science.1232033

Jinek M, Chylinski K, Fonfara I, et al (2012) A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science* 337:816–21. doi: 10.1126/science.1225829

Cong L, Ran FA, Cox D, et al (2013) Multiplex genome engineering using CRISPR/Cas systems. *Science* 339:819–23. doi: 10.1126/science.1231143

Singh P, Schimenti JC, Bolcun-Filas E (2014) A Mouse Geneticist's Practical Guide to CRISPR Applications. *Genetics* genetics.114.169771–. doi: 10.1534/genetics.114.169771

Brandl C, Ortiz O, Röttig B, et al (2015) Creation of targeted genomic deletions using TALEN or CRISPR/Cas nuclease pairs in one-cell mouse embryos. *FEBS Open Bio* 5:26–35. doi: 10.1016/j.fob.2014.11.009

Zhou J, Wang J, Shen B, et al (2014) Dual sgRNAs facilitate CRISPR/Cas9 mediated mouse genome targeting. *FEBS J*. doi: 10.1111/febs.12735

Kraft K, Geuer S, Will AJ, et al (2015) Deletions, Inversions, Duplications: Engineering of Structural Variants using CRISPR/Cas in Mice. *Cell Rep*. doi: 10.1016/j.celrep.2015.01.016

Shen B, Zhang J, Wu H, et al (2013) Generation of gene-modified mice via Cas9/RNA-mediated gene targeting. *Cell Res* 23:720–3. doi: 10.1038/cr.2013.46

Wang H, Yang H, Shivalila CS, et al (2013) One-step generation of mice carrying mutations in multiple genes by CRISPR/Cas-mediated genome engineering. *Cell* 153:910–8. doi: 10.1016/j.cell.2013.04.025

Yang H, Wang H, Shivalila CS, et al (2013) One-Step Generation of Mice Carrying Reporter and Conditional Alleles by CRISPR/Cas-Mediated Genome Engineering. *Cell* 154:1370–1379. doi: 10.1016/j.cell.2013.08.022

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