

MGPgenotyping@sanger.ac.uk www.sanger.ac.uk

Gene: Zkscan14

Colony prefix: MDAY

ESC clone ID: EPD0147\_2\_E01

Allele: Zkscan14<sup>tm1a(EUCOMM)Wtsi</sup>

WT PCR

Allele type: Knockout First, Reporter-tagged insertion with conditional potential

# Allele information:

Further information about the allele can be found on the 'International Mouse Phenotyping Consortium' (IMPC) web site at http://www.mousephenotype.org/martsearch\_ikmc\_project/martsearch/ikmc\_project/24619 Details on how to determine the floxed exon can be found at http://www.i-dcc.org/kb/entry/21/

# Mouse QC information



Promoter-Driven Cassette shown for illustrative purposes

Southern Blot	na	TV Backbone Assay	pass	5' LR-PCR	na
Loss of WT Allele (LOA) qPCR	pass	Homozygous Loss of WT Allele (LOA) SR-PCR	pass	Neo Count (qPCR)	pass
LacZ SR-PCR	pass	5' Cassette Integrity	pass	Neo SR-PCR	na
Mutant Specific SR- PCR	pass	LoxP Confirmation	pass	3' LR-PCR	na
Genotyping Comment					

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## Southern blot confirmation:

Southern blots are not routinely performed at the Sanger Institute due to throughput constraints. A southern blot experiment design tool can be found on the IMPC web site at http://www.mousephenotype.org/martsearch\_ikmc\_project/martsearch/ikmc\_project/24619

# Links to information and frequently asked questions about the EUCOMM/KOMP alleles and MGP projects

General targeting strategies: http://www.mousephenotype.org/martsearch\_ikmc\_project/about/targeting-strategies

MGP mouse phenotype data: http://www.sanger.ac.uk/mouseportal/

IKMC allele types: http://www.i-dcc.org/kb/entry/89/

MGP mouse quality control tests : http://www.i-dcc.org/kb/25/

Allele conversion guide - genotyping tm1b, tm1c and tm1d mice: http://www.i-dcc.org/kb/entry/105/

How the "critical" exon is decided: http://www.i-dcc.org/kb/entry/102/

# **Genotyping Information**

### Genotyping by end-point PCR

These mice may be genotyped through a combination of separate PCR reactions that detect the cassette, the gene-specific wild type allele, and a mutant allele-specific short range PCR. Interpretation of the consolidated results produces the genotype of the mice.

For example: cassette positive, mutant positive, wild type positive = heterozygous.

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# PCRs primer pairs and expected size bands

Assay Type	Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Standard PCR	Wildtype	Zkscan14_48468_F	Zkscan14_48468_R	345
Standard PCR	Mutant	Zkscan14_48468_F	CAS_R1_Term	116
Standard PCR	Cassette	LacZ_2_small_F	LacZ_2_small_R	108

## **Primer sequences**

Primer Name	Primer Sequence (5' > 3')
CAS_R1_Term	TCGTGGTATCGTTATGCGCC
LacZ_2_small_F	ATCACGACGCGCTGTATC
LacZ_2_small_R	ACATCGGGCAAATAATATCG
Zkscan14_48468_F	TGTGCAGGAGTGTGGGGTAG
Zkscan14_48468_R	ATCCCATGAATGTGGGCTG

### **Reaction setup**

Reagent	μί
DNA (~50-100 ng)	1
10x Buffer	2
MgCl2 (50 mM)	0.6
Platinum Taq (Invitrogen)	0.2
dNTPs (100 mM)	0.2
Primer 1 (10 M)	0.4
Primer 2 (10 M)	0.4
ddH20	15.2
Total	20

# Amplification conditions

Step	Conditions	Time
1	94°C	5 min
2	94°C	30 sec
3	58°C	30 sec
4	72°C	45 sec
5	Go to '2' + 34 cycles	-
6	72°C	5 min
7	12°C	forever

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#### Genotyping by loss of WT allele qPCR Assay (gene-specific assay)

The wild type loss of allele (LoA) qPCR assay uses a hydrolysis probe assay (for example Applied Biosystems TaqMan® technology) to determine the copy number of the wild type allele in a sample. Homozygotes will show no amplification, heterozygotes one copy and wild type mice will show two copies when compared to a wild type control.

The number of copies of the Zkscan14 allele can be detected using a FAM-labelled custom qPCR TaqMan® assay. These are multiplexed with a VIC® labelled endogenous control assay (for example TaqMan® Copy Number Reference Assay, Mouse, Tfrc; Applied Biosystems part #4458366). Reference DNA controls of known genotypes should also be included to facilitate correct analysis.

#### Primers for LoA qPCR assay

Primer type	Assay Name	Forward Primer Seq.	Reverse Primer Seq.	Probe Primer Seq.
LoA	Zkscan14_WT	AGCTGGTGATGCCAACATATGAA	GTATGCACCTTCACCCAATGCTA	CCAATGCCAGTTCAGCC

#### **Reaction setup**

Reaction setup and amplification conditions are the same as those used for the neo cassette qPCR assay.

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#### **Relevant publications**

Ryder, E., Doe, B., Gleeson, D., Houghton, R., Dalvi, P., Grau, E., Ramirez-Solis, R. (2013). Rapid conversion of EUCOMM/KOMP-CSD alleles in mouse embryos using a cell-permeable Cre recombinase. Transgenic research. 23(1), 177–185.

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White, J.K., Gerdin, A.-K., Karp, N.A., Ryder, E., Buljan, M., Bussell, J.N., Salisbury, J., Clare, S., Ingham, N.J., Podrini, C., et al. (2013). Genome-wide Generation and Systematic Phenotyping of Knockout Mice Reveals New Roles for Many Genes. Cell 154, 452–464.

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Bradley, A., Anastassiadis, K., Ayadi, A., Battey, J.F., Bell, C., Birling, M.-C., Bottomley, J., Brown, S.D., Bürger, A., Bult, C.J., et al. (2012). The mammalian gene function resource: the international knockout mouse consortium. Mamm Genome 23, 580–586.

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Liang, Q., Conte, N., Skarnes, W.C., and Bradley, A. (2008). Extensive genomic copy number variation in embryonic stem cells. Proc Natl Acad Sci U S A 105, 17453–17456.

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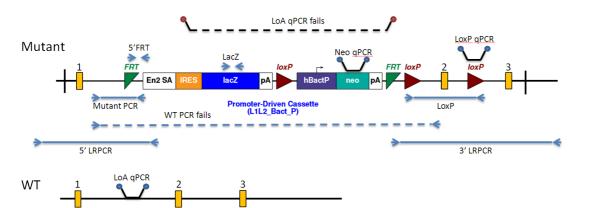
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