

MGPgenotyping@sanger.ac.uk www.sanger.ac.uk

Gene: Tk1

Colony prefix: MTKB

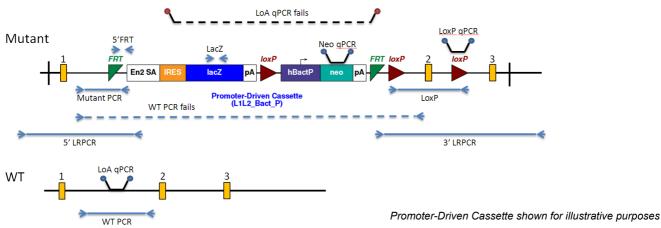
ESC clone ID: Allele: Tk1<sup>tm1Brd</sup>

Allele type: Knockout First, Reporter-tagged insertion with conditional potential

Allele information:

Details on how to determine the floxed exon can be found at http://www.knockoutmouse.org/kb/entry/21/

#### Mouse QC information



Southern Blot

TV Backbone Assay

5' LR-PCR

Loss of WT Allele
(LOA) qPCR

Homozygous Loss of WT
Allele (LOA) SR-PCR

Neo Count (qPCR)

Neo SR-PCR

Neo SR-PCR

Mutant Specific SR-PCR

LoxP Confirmation

Genotyping Comment

This technical data sheet and information ("Datasheet") is supplied by Genome Research Limited ("GRL").



MGPgenotyping@sanger.ac.uk www.sanger.ac.uk

#### Southern blot confirmation:

Southern blots are not routinely performed at the Sanger Institute due to throughput constraints.

# Links to information and frequently asked questions about the EUCOMM/KOMP alleles and MGP projects

General targeting strategies:

http://www.mousephenotype.org/martsearch\_ikmc\_project/about/targeting-strategies

MGP mouse phenotype data:

http://www.sanger.ac.uk/mouseportal/

IKMC allele types:

http://www.i-dcc.org/kb/entry/89/

MGP mouse quality control tests:

http://www.i-dcc.org/kb/25/

Allele conversion guide - genotyping tm1b, tm1c and tm1d mice:

http://www.i-dcc.org/kb/entry/105/

How the "critical" exon is decided:

http://www.i-dcc.org/kb/entry/102/

#### **Genotyping Information**

#### Genotyping by end-point PCR

These mice may be genotyped through a combination of separate PCR reactions that detect the cassette, the gene-specific wild type allele, and a mutant allele-specific short range PCR. Interpretation of the consolidated results produces the genotype of the mice

For example: cassette positive, mutant positive, wild type positive = heterozygous.

This technical data sheet and information ("Datasheet") is supplied by Genome Research Limited ("GRL").



MGPgenotyping@sanger.ac.uk www.sanger.ac.uk

### PCRs primer pairs and expected size bands

Assay Type	Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Standard PCR	Wildtype	Tk1_F	Tk1_WT_R	66
Standard PCR	Mutant	Tk1_F	Tk1_Mut_R	74

## **Primer sequences**

Primer Name	Primer Sequence (5' > 3')	
Tk1_F	CCCTTAAATGACTTCTCTAGGT	
Tk1_Mut_R	GGCCCAAAGCTTCATTAACCTTTAC	
Tk1_WT_R	CCATTCATTACCTTTTCCCT	

## **Reaction setup**

Reagent	μΙ
DNA (~50-100 ng)	1
10x Buffer	2
MgCl2 (50 mM)	0.6
Platinum Taq (Invitrogen)	0.2
dNTPs (100 mM)	0.2
Primer 1 (10 µM)	0.4
Primer 2 (10 µM)	0.4
ddH20	15.2
Total	20

## **Amplification conditions**

Step	Conditions	Time
1	94°C	5 min
2	94°C	30 sec
3	58°C	30 sec
4	72°C	45 sec
5	Go to '2' + 34 cycles	-
6	72°C	5 min
7	12°C	forever

This technical data sheet and information ("Datasheet") is supplied by Genome Research Limited ("GRL").



MGPgenotyping@sanger.ac.uk www.sanger.ac.uk

#### Relevant publications

Ryder, E., Doe, B., Gleeson, D., Houghton, R., Dalvi, P., Grau, E., Ramirez-Solis, R. (2013). Rapid conversion of EUCOMM/KOMP-CSD alleles in mouse embryos using a cell-permeable Cre recombinase. Transgenic research. 23(1), 177–185.

Ryder, E., Gleeson, D., Sethi, D., Vyas, S., Miklejewska, E., Dalvi, P., Habib, B., Cook, R., Hardy, M., Jhaveri, K., et al. (2013). Molecular Characterization of Mutant Mouse Strains Generated from the EUCOMM/KOMP-CSD ES Cell Resource. Mamm. Genome, 24, 286–294.

White, J.K., Gerdin, A.-K., Karp, N.A., Ryder, E., Buljan, M., Bussell, J.N., Salisbury, J., Clare, S., Ingham, N.J., Podrini, C., et al. (2013). Genome-wide Generation and Systematic Phenotyping of Knockout Mice Reveals New Roles for Many Genes. Cell 154, 452–464.

Ryder, E., Wong, K., Gleeson, D., Keane, T.M., Sethi, D., Vyas, S., Wardle-Jones, H., Bussell, J.N., Houghton, R., Salisbury, J., et al. (2013). Genomic analysis of a novel spontaneous albino C57BL/6N mouse strain. Genesis 51, 523–528.

Bradley, A., Anastassiadis, K., Ayadi, A., Battey, J.F., Bell, C., Birling, M.-C., Bottomley, J., Brown, S.D., Bürger, A., Bult, C.J., et al. (2012). The mammalian gene function resource: the international knockout mouse consortium. Mamm Genome 23, 580–586.

Birling, M.-C., Dierich, A., Jacquot, S., Hérault, Y., and Pavlovic, G. (2011). Highly-efficient, fluorescent, locus directed Cre and flpo deleter mice on a pure C57BL/6N genetic background. Genesis.

Skarnes, W.C., Rosen, B., West, A.P., Koutsourakis, M., Bushell, W., Iyer, V., Mujica, A.O., Thomas, M., Harrow, J., Cox, T., et al. (2011). A conditional knockout resource for the genome-wide study of mouse gene function. Nature 474, 337–342.

Pettitt, S.J., Liang, Q., Rairdan, X.Y., Moran, J.L., Prosser, H.M., Beier, D.R., Lloyd, K.C., Bradley, A., and Skarnes, W.C. (2009). Agouti C57BL/6N embryonic stem cells for mouse genetic resources. Nat Methods 6, 493–495.

Liang, Q., Conte, N., Skarnes, W.C., and Bradley, A. (2008). Extensive genomic copy number variation in embryonic stem cells. Proc Natl Acad Sci U S A 105, 17453–17456.

Farley, F.W., Soriano, P., Steffen, L.S., and Dymecki, S.M. (2000). Widespread recombinase expression using FLPeR (flipper) mice. Genesis 28, 106–110.

This technical data sheet and information ("Datasheet") is supplied by Genome Research Limited ("GRL").