Transcriptomic studies on host-parasite interactions in *Schistosoma mansoni* intramammalian stages

University of Cambridge Pembroke College



A thesis submitted for the degree of Doctor of Philosophy

Arporn Wangwiwatsin

The Wellcome Trust Sanger Institute

March 2017

Declaration

The work presented in this thesis was carried out at the Wellcome Trust Sanger Institute (Hinxton) between October 2012 and March 2017. This dissertation is the result of my own work – contributions from collaborations are clearly referenced. No part of this dissertation has been or is being submitted for any qualification in any other university. This thesis does not exceed the word limit established by the Biology Degree Committee.

Acknowledgements

First of all I would like to pass a massive gratitude to my supervisor, Matt, for giving me this opportunity, and for the understanding, patience, guidance, and support throughout. Thanks to everyone in the Parasite Genomics team including past and current members. Thanks for all the ideas, the fruitful discussions, and all the fun in team outing. Many people also helped read this thesis: Matt, Gabriel, Patrick, Adam, Kate, Anna, Avril, and Shona. Thank you to my thesis examiners for their contributions to the improvement of this thesis. Thank you Christina and Annabel from the Graduate Office for all their help with all questions I can ever ask. Thank you to Marc who has made every printing request possible.

Many people have been amazingly supportive and helpful throughout my work at the Sanger Institute, sharing their knowledge, helping make my experiment endeavour come true and many have become good friends. Thanks CGaP, Sally Forrest, Christ Kirton, Emma Goss, Theresa Feltwell, Liam Prestwood, Laura Wood, James Hewinson, Gavin Wright, Paul Kellam, Mercedes Pardo, Jyoti Choudhari, Lira Mamanova, Rachael Wash, Sarah Smith, Sally Linsdell, Derek Pickard, Dave Goulding Simon Clare, Cordelia Brandt, Colin Barker, Carol Smee, Dan Mead, Theodoros Roumeliotis (especially for his help wth using the IPA), and the Sanger librarians.

Thanks for all support and joy beyond the work at Sanger Institute including Pembroke College, Public engagement teams at the Sanger and Cambridge, the CHaOS Society, Cambridge Thai Society, University Counselling Service, Polly Brown, family, and the husband.

It has been a wonderful experience.

Summary

The life cycle of the parasitic flatworm *Schistosoma mansoni* is split between snail and human hosts. In humans, it lives in the bloodstream and can survive for over 10 years, during which time, interactions with the host are essential for its survival. Starting the infection, cercariae penetrate human skin and become schistosomules which enter blood vessels. The schistosomules follow blood circulation to the lung and the liver where they develop into adults which migrate to the mesenteric venules for egg-laying. The association with certain host tissues and egg-laying site may involve interactions and adaptations. Moreover, the parasite needs to evade or modulate host immune responses and to optimise its acquisition of host metabolites. Molecular mechanisms involved in these interactions are not completely understood.

To gain deeper understanding into the interactions of *S. mansoni* and its mammalian host, I sought to understand what biological processes are required for successful intramammalian infection; specifically, what guides the tissue tropism of the parasite and what roles do the host tissues play in the infection? In the first part, parasite transcriptomes were produced from *S. mansoni* obtained from experimentally infected mice. The dataset includes novel transcriptomic profile of the lung stage, and covers developmental and egg-laying stages. In the next two parts, co-culture experiments were set up using mechanically transformed schistosomules and cells derived from human tissues. Transcriptomic profiles of the co-cultured schistosomules were investigated in the second part of this thesis; and transcriptomic profiles of the co-cultured human cells were investigated in the final part. The outputs of this thesis provide new insights into the infection biology, provide a large data resource for the research communiy, and propose avenues for further investigation and characterisation of interaction mechanisms.

Abbreviations

BLAST Basic Local Alignment Search Tool

C1 Complement component 1

CFH Complement factor H
CFI Complement factor I

DAF Decay-accelerating factor

DMEM Dulbecco's Modified Eagle's Medium

DMT Divalent metal transporter

ES Excretory/secretory products

ESTs Express Sequence Tags

FPKM Fragments Per Kilobase of transcript per Million mapped reads

GO Gene Ontology

GPCR G-protein coupled receptor

ICAM1 Intercellular adhesion molecule 1

IL-2 Interleukin-2

IPA Ingenuity Pathways Analysis

KEGG Kyoto Encyclopedia of Genes and Genomes

LDL Low density lipoprotein

Log₂FC Log₂ fold change

LPS Lipopolysaccharide

MEG Micro-exon gene

NPC2 Niemann Pick type C2 protein

PCA Principal component analysis

PDB Protein Data Bank

RNA-seq RNA-sequencing

ROS Reactive oxygen species

SELE Selectin E

TAL Tegumental-allergen-like

TGF-β Transforming growth factor-beta

TNF-α Tumour-necrosis factor-alpha

VCAM1 Vascular cell adhesion molecule

WTSI Wellcome Trust Sanger Institute

Table of Contents

Chapter 1

Intro	duction	1
1.1	Overview	1
1.2	Introduction to schistosomes	2
1.2	2.1 Schistosome life cycle	2
1.2	2.2 Schistosomiasis	5
1.3	S. mansoni life in bloodstream	7
1.3	Host-parasite interactions: the interfaces	7
1.3	Motility, behaviour, and homeostasis: the nervous system	11
1.3	3.3 Migration route and development	12
1.3	Metabolic requirements and acquisition	17
1.3	3.5 Immune evasion	22
1.3	3.6 Section summary	30
1.4	S. mansoni genome and transcriptome	30
1.4	I.1 Genome and gene annotation	30
1.4	1.2 Transcriptomes	33
1.5	This thesis	34
Chap	oter 2	
Mate	erials and methods	35
2.1	Parasite materials	35
2.1	.1 Maintenance of the S. mansoni life cycle	35
2.1		
2.1	.3 Mouse maintenance	37
2.2	Molecular methods	37

2.2.1	RNA extraction from parasites	37
2.2.2	RNA extraction from human cells	38
2.2.3	Measurements of RNA concentration and purity	38
2.2.4	Library production and sequencing	39
2.2.5	Overall QC of sequencing outcome	40
2.3 Da	ta analysis	41
2.3.1	Mapping and quantifying read counts	41
2.3.2	Read count and differential expression analysis	42
2.3.3	Genes clustering by timecourse expression profile	42
2.3.4	GO term enrichment	43
2.3.5	Pathway enrichment and pathway network	43
2.3.6	Pathway comparison between cell types	44
2.3.7	Protein structural prediction	45
2.3.8	Protein domains and motif search	45
2.3.9	Gene phylogenetic tree	45
2.3.10	Artemis and BamView	45
2.3.11	Seaview sequence alignment	46
	Seaview sequence alignmentta presentation	
2.4 Da Chapter	ta presentation	46
2.4 Da Chapter S. manso	ta presentation	46
2.4 Da Chapter S. manso	3 oni in experimentally infected mice	4647
2.4 Da Chapter S. manso 3.1 Inter	3 oni in experimentally infected mice	4647
2.4 Da Chapter S. manso 3.1 Into	3 oni in experimentally infected mice roduction	464747
2.4 Da Chapter S. manso 3.1 Into 3.1.1 3.1.2	a presentation oni in experimentally infected mice roduction Overview Host-parasite interactions in intra-mammalian stages	46474747
2.4 Da Chapter S. manso 3.1 Into 3.1.1 3.1.2 3.1.3	a presentation oni in experimentally infected mice roduction Overview Host-parasite interactions in intra-mammalian stages Progress in transcriptomic and genomic approaches	4647474749
2.4 Da Chapter S. manso 3.1 Inti 3.1.1 3.1.2 3.1.3 3.1.4 3.1.5	ani in experimentally infected mice roduction Overview	464747474949
2.4 Da Chapter S. manso 3.1 Inti 3.1.1 3.1.2 3.1.3 3.1.4 3.1.5	ani in experimentally infected mice Toduction	464747494950

3.2.3	RNA extraction and library preparation	54
3.2.4	Morphology score	54
3.3 Re	sults	55
3.3.1	Worm morphology	55
3.3.2	RNA quantity and quality	58
3.3.3	Sequencing yields	58
3.3.4	Sequencing data overall profiles	60
3.3.5	Lung stage	64
3.3.6	Liver stages	76
3.3.7	Adult stages	83
3.4 Dis	scussion	94
3.4.1	Overview	94
3.4.2	Potential effect of collection procedures	94
3.4.3	Lung stage signalling	95
3.4.4	Lung stage immune evasion	96
3.4.5	Complement factor H	96
3.4.6	Complement factor I	97
3.4.7	Immunomodulation in liver stages	97
3.4.8	Implication for intervention	98
3.4.9	Expected changes in liver and adult stages	98
3.4.10	Liver localisation	98
3.4.11	Mesenteric migration	99
3.4.12	Micro-exon genes	101
3.4.13	Other ways to use this data	101
3.4.14	Summary	101
Chapter	4	
Chapter	4	
S. manso	oni in vitro culture with cell lines	103
4.1 Int	roduction	103
4.1.1	Overview	103

4	4.1.2	S. mansoni and host tissues	103
4	4.1.3	Effects of host environments on <i>S. mansoni</i>	104
4	4.1.4	Aims and approaches	105
4	4.1.5	Chapter outline	105
4.2	2 Met	thods	106
4	4.2.1	Experimental design	106
4	4.2.2	Preparing human cells: maintaining stock cells	108
4	4.2.3	Preparing schistosomules: media for schistosomules and	
		for the co-culture	110
4	4.2.4	Preparing schistosomules: transforming cercariae into schistosomule	es 110
4	4.2.5	Co-culture	111
4	4.2.6	Sample collection	112
4.3	B Res	ults	113
4	4.3.1	Mycoplasma test	113
4	4.3.2	Worm morphology	114
4	4.3.3	RNA quantity and quality	116
4	4.3.4	Overall profiles of transcriptomes	116
4	4.3.5	Overview of gene expression	119
4	4.3.6	Schistosomules in co-cultured environment: generic responses	125
4	4.3.7	Schistosomule adaptation to HEPG2 environment	133
4	4.3.8	Schistosomule adaptation to HUVEC environment	140
4.4	4 Disc	cussion	146
4	4.4.1	Overview	146
4	4.4.2	In vitro schistosomules and effect of culture methods	147
4	4.4.3	Generic responses	149
4	4.4.4	HEPG2-specific responses	150
4	4.4.5	HUVEC-specific responses	151
4	4.4.6	Summary	152

Chapter 5

Transcri	ptomes	of	cell	lines
11 4113611	Ptomes	$\mathbf{O}_{\mathbf{I}}$	CCII	111103

expose	d to schistosomules in vitro	. 155
5.1 lı	ntroduction	155
5.1.1	Overview	155
5.1.2	Host responses in <i>S. mansoni</i> infections	155
5.1.3	Host tissue responding to <i>S. mansoni</i>	156
5.1.4	Aims and approaches	156
5.1.5	Outline	157
5.2 N	Methods	157
5.3 F	Results	158
5.3.1	RNA quantity and quality	158
5.3.2	HUVEC and endothelial cell surface markers	159
5.3.3	HEPG2 and liver hepatocytes	161
5.3.4	Overall profiles of transcriptomes	161
5.3.5	Differential expression between co-cultured vs. worm-free cells	163
5.3.6	Pathways between cell types	183
5.4	Discussion	187
5.4.1	Overview	187
5.4.2	On in vitro-adapted cells	188
5.4.3	On aged media	189
5.4.4	Biological functions affected by parasite co-culture	189
5.4.5	On in vivo validation	193
5.4.6	Summary	193
Chapte	r 6	
Discuss	ion and conclusion	195
6.1 F	indings and interpretations	196

6.1	.1	Interactions with host defences	196
6.1	.2	Host reparation	198
6.1	.3	Responses to environments	198
6.2	Lim	nitations	201
6.3	Со	ncluding remarks	202
Chap Refer		7 ces	203
Chap	ter	8	
Appe	ndi	ces	235

List of Figures

Figure 1.1 Life cycle of Schistosoma spp
Figure 1.2 The schistosome tegument
Figure 1.3 Coagulation and complement cascade
Figure 1.4 Coagulation pathway
Figure 2.1 Transfer of snail eggs using "The Octopus"
Figure 2.2 RNA qualities assessed from Agilent Bioanalyzer electropherograms39
Figure 3.1 Experiment layout
Figure 3.2 Diagram used for morphology scoring
Figure 3.3 Morphology of in vivo S. mansoni
Figure 3.4 PCA plots from total reads and down-sized reads
Figure 3.5 Similarity between 96 clusters
Figure 3.6 Clusters of genes based on timecourse expression pattern
Figure 3.7 Clusters of genes based on timecourse expression pattern, with fixed y-axes 63
Figure 3.8 Volcano plots of all consecutive pairwise comparison
Figure 3.9 GO enrichment of genes up-regulated in day-6 compared to day-13 schistosomules
Figure 3.10 MEGs differentially expressed between day-6 and day-13 schistosomules 69
Figure 3.11 Predicted structure of Smp_182770 aligned with structure of human CFH 73
Figure 3.12 Homologous relationship of Smp_182770 and alignment of RNA-seq reads to
genomic locations
Figure 3.13 Clusters of genes with high expression in liver stages and GO term enrichment 78
Figure 3.14 GO enrichment of genes up-regulated in day-13 compared to day-6
schistosomules
Figure 3.15 GO enrichment of genes up-regulated in day-35 compared to day-28 worms 86
Figure 3.16 GO enrichment of genes up-regulated in day-28 compared to day-35 worms 87
Figure 3.17 Expression profile of GPCRs up-regulated in day-28 compared to day-35 worms
Figure 3.18 Log ₂ FC of genes in cluster 40 in response to pairing
Figure 4.1 Experiment set up
Figure 4.2 Gel image of PCR product from <i>Mycoplasma</i> tests
Figure 4.3 Example images from each time point of each co-culture condition
Figure 4.4 PCA of transcriptomes from in vitro and in vivo datasets
Figure 4.5 PCA of transcriptomes from all <i>in vitro</i> parasites and <i>in vivo</i> lung schistosomules
118

Figure 4.6 PCA of transcriptomes from all <i>in vitro</i> schistosomules	119
Figure 4.7 MA plots of pairwise comparisons between co-cultured and control schistosome	mules
	120
Figure 4.8 Differentially expressed genes unique and common to each pairwise comparis	son
	121
Figure 4.9 Differentially expressed genes in day-17 schistosomules comparing unique ar	ıd
common changes between cell types	122
Figure 4.10 Enriched GO terms in genes up-regulated in day-17 co-cultured schistosomu	ıles
	123
Figure 4.11 Enriched GO terms in genes down-regulated in day-17 co-cultured	
schistosomules	124
Figure 4.12 Generic responses in schistosomules	126
Figure 4.13 Homologous relationship of Smp_067800	127
Figure 4.14 Genomic region of Smp_067800 and alignment of RNA-seq reads	129
Figure 4.15 Homologous relationship of Smp_052880	131
Figure 4.16 Smp_052880 genomic region and amino acid sequence alignment with other	ſ
BLASTP hits	132
Figure 4.17 Volcano plot of pairwise comparison between HEPG2 vs. non-HEPG2	
schistosomules	134
Figure 4.18 Enriched GO terms in genes up-regulated in HEPG2 compared to non-HEPG	3 2
schistosomules	135
Figure 4.19 Enriched GO terms in genes down-regulated in HEPG2 compared to non-HE	EPG2
schistosomules	135
Figure 4.20 Volcano plot of pairwise comparison between HUVEC vs. non-HUVEC	
schistosomules	141
Figure 4.21 Smp_123920 homologous relationship and structural prediction	143
Figure 4.22 Expression profiles of Smp_123920 in vivo and in vitro	144
Figure 4.23 Smp_016490 domains and expression profiles	145
Figure 4.24 Smp_052880 in vivo expression profile	150
Figure 5.1 260/230 ratio of RNA extracted from human cells and its relationship with the	Э
RNA concentration	159
Figure 5.2 Expression levels of endothelial cell surface markers in worm-free HUVEC	160
Figure 5.3 PCA from transcriptomes of all cell types, and separate for each cell type	162
Figure 5.4 Volcano plots for pairwise comparison in each cell type between co-cultured	vs.
worm-free conditions	164
Figure 5.5 Expression profiles of SELE VCAM1, and ICAM1	165

Figure 5.6 Expression of endothelial adhesion molecules between co-cultured vs. worm-free
HUVEC
Figure 5.7 GO enrichment of genes differentially expressed in co-cultured HUVEC compared
to worm-free HUVEC
Figure 5.8 Pathway enrichment of genes differentially expressed in co-cultured HUVEC
compared to worm-free HUVEC
Figure 5.9 Coagulation-related genes up-regulated in co-cultured HUVEC
Figure 5.10 GO enrichment of genes differentially expressed in co-cultured HEPG2 compared
to worm-free HEPG2
Figure 5.11 Pathway enrichment of genes differentially expressed in co-cultured HEPG2
compared to worm-free HEPG2
Figure 5.12 GO enrichment of genes differentially expressed in co-cultured GripTite
compared to worm-free GripTite
Figure 5.13 Pathway enrichment of genes differentially expressed in co-cultured GripTite
compared to worm-free GripTite
Figure 5.14 Immune responses-related genes up-regulated in co-cultured GripTite
Figure 5.15 Log ₂ FC of genes in extracellular matrix organisation pathways
Figure 5.16 Log ₂ FC of genes in coagulation and complement cascade

List of Tables

Table 3.1 Average of total read counts
Table 3.2 Genes in cluster 8 (highly expressed in lung stage)
Table 3.3 Enriched GO terms of genes down-regulated after lung stage
Table 3.4 Top 20 genes up-regulated in day-6 compared to day-13 schistosomules 67
Table 3.5 Genes in cluster 72
Table 3.6 Top 20 genes up-regulated in day-13 compared to day-6 schistosomules
Table 3.7 Enriched GO terms (biological process) of genes up-regulated in day-21 compared
to day-28 worms
Table 3.8 Top 20 genes up-regulated in day-28 compared to day-21 worms
Table 3.9 Enriched GO terms (biological process) of genes up-regulated in day-28 compared
to day-21 worms
to day-21 worms
Table 3.10 Genes in cluster 40

List of Appendices

Appendix A: Aquarium water, 10X	235
Appendix B: Reference genomes used by NPG QC	236
Appendix C: Versions of R packages used in data analysis	238
Appendix D: Basch media components	239
Appendix E: S. mansoni genetrees downloaded from WormBase ParaSite release 9	240
Appendix F: Orthologues of Smp_041700 downloaded from WormBase ParaSite release	se 8
	251
Appendix G: List of endothelial cell surface marker from Durr et al., 2004	252
Appendix H: List of supplementary tables	255