A Study of Molecular Synergy and Clonal Evolution in Haematopoietic Malignancies

Carolyn Suzanne Grove

Emmanuel College, Cambridge August 2014

This dissertation is submitted for the degree of Doctor of Philosophy

Declaration

This dissertation is the result of work undertaken in the laboratory of Dr George Vassiliou at the Wellcome Trust Sanger Institute. The dissertation is the result of my own work, except where specific reference is made to the work of others. Where data is the result of collaboration with others, this is clearly stated as such in the text. This work has not previously been submitted for any other degree or qualification. The text of this dissertation, excluding tables, figures and references, does not exceed 60 000 words.

Carolyn Grove August 2014

Acknowledgements

I would like to thank my supervisor, George Vassiliou, who has been an enthusiastic and encouraging mentor, role model and friend. I consider it a privilege to have been his first PhD student and to be one of the early members of his laboratory, which is a dynamic, stimulating and pleasurable place to work.

I would also like to thank my co-supervisor Martin Turner, the other members of my thesis committee, Peter Campbell and Allan Bradley, and my 'unofficial' supervisor Roland Rad. All have given me considerable time, encouragement and support and I have benefited from their diverse expertise and willingness to help me to develop my skills.

This thesis work was completed with considerable support from other members of team 163 at the Wellcome Trust Sanger Institute (WTSI). I would particularly like to thank Jon Cooper, Meg Byrne and Sarah Paterson, all of whom generously assisted with the routine aspects of this work, in particular mouse genotyping, necropsies, blood analysis, tissue storage and DNA extraction. Jon taught me many laboratory skills and has been an enthusiastic helper and good friend. Meg looked after my mouse colonies during my period of maternity leave and Sarah kindly assisted me during two summer vacations, while also studying for her medical degree at Glasgow University. I would also like to thank the other members of team 163, all of whom provided me with useful insights and helped to generate such an enjoyable atmosphere in which to work.

There are several people who helped with particular aspects of this work, which are recognised throughout the text, but I would also like to acknowledge them here. Gary Hoffman, Iraad Bronner, Kosuke Yusa, Nicla Manes, Stephen Rice, Hannes Ponstingl, Ignacio Varela, David Wedge and Rachael Bashford-Rogers have all been tremendously supportive. Also, this work would not have been possible without the generous support of Leukaemia Lymphoma Research. I will always be grateful to this charity for the wonderful opportunity they have given me.

Finally, I would like to thank my family, in particular my husband Jonathan Francis, who has assisted in so many ways. This dissertation is much more visually attractive than it would have been without his help in formatting the many tables. It would not exist at all, without the strength, love and support that he has given me. I would also like to thank my mother-in-law Annette, who has been a source of inspiration in her resilience, especially over recent months. In addition I would like to thank Catherine and Malcolm, my sister- and father-in-law who supported me through so much and who shall always be remembered for their generosity of spirit. I also want to thank my parents, who have always been there for me and have willingly supported me in every endeavour. Finally, to Matthew, who came into the world during the course of this work and has kept a smile on my face, even during the toughest periods.

Abstract

Haematopoeitic malignancies evolve through the serial selection of cells with a growth advantage, in a multi-step process akin to natural selection. Transposon insertional mutagenesis (IM) is a powerful approach for the identification and validation of cancer driver mutations and compliments human sequencing efforts. This technology has not previously been applied to study tumour evolution, nor has the sub-clonal architecture of transposon driven tumours been carefully investigated.

In the first part of this work I have investigated the timing and pattern of acquisition of mutations in NPM1-mutant acute myeloid leukaemia (AML). NPM1 mutations are found in around 30% of cases of AML and are thought to be critical events in leukaemogenesis. First, I present the detailed study of an informative human case of CMML evolving to AML and discuss the implications for clonal evolution and leukaemic transformation. Subsequently, I describe the investigation of an IM mouse model of Npm1-mutant AML in which the timing and order of acquisition of transposon integrations was characterised using pre-leukaemic blood samples. The driver status and co-occurrence of integrations was also investigated in serial transplant experiments. Transposon mobilisation continued throughout leukaemia evolution, but this data suggests that only a minority of integrations behave as 'driver' mutations in this context. Although some of these 'drivers' were detectable several weeks earlier, the onset of leukaemia was sudden and occurred without antecedent abnormalities in blood count parameters. Transplant experiments demonstrated that multiple distinct clones with different transposon integrations were present within the primary tumour cell population.

In the final part of this dissertation I present the findings of two mouse models in which *piggyBac* (*PB*) IM is targeted to the mature B cell compartment for cancer gene discovery. Both models were based on the published *Vk*MYC* mice, which were reported to develop highly penetrant plasma cell malignancies recapitulating the major features of human multiple myeloma. In one model, the *PB* transposase replaced the *MYC* transgene in the *Vk*MYC* construct. In the second, *MYC* and *PB* were co-expressed from the same cistron, in order to identify genes co-operating with *MYC* in oncogenesis. IM mice had a significantly reduced survival largely due to the development of mature B cell lymphomas; although plasma cell malignancies were not a feature. Mapping and common integration site analysis of transposon

A Study of Molecular Synergy and Clonal Evolution in Haematopoietic Malignancies

insertions identified several recurrent integrations in known (e.g. *Bcl6*) and novel (e.g. *Rreb1*) lymphoma-associated genes.

Contents

Introduction

1.1 Cancer as an evolutionary process	.9
1.2 AML as an exemplar of clonal evolution	13
1.2.1 How many driver mutations are required for leukaemogenesis?	13
1.2.2 Genotype Phenotype Correlations and Myeloid Malignancy	16
1.2.3 Linear Versus Branching Evolution and Clonal Hierarchy	17
1.2.4 The Timeframe for AML Evolution	20
1.2.5 Initiating Mutations and Order of Acquisition	22
1.3 AML with mutated NPM1	24
1.4 Transposons as tools for gene discovery in the study of cancer	26
1.5 Using transposon insertional mutagenesis to study the molecular pathogenesis of AML	31
1.6 Transposon insertional mutagenesis for cancer gene discovery in mature B cell malignancies	32
1.6.1 Normal B cell development	32
1.6.2 Correlation of lymphoma phenotypes with normal B cell development	33
1.6.3 Modelling Mature B cell Neoplasms in the Mouse	34
1.6.4 Targeting insertional mutagenesis to the mature B cell compartment	36
1.7 AIMS	36

Methods

2.1 Sequencing of human leukaemia samples	. 38
2.1.1Exome Sequencing and genomic alignment	.38
2.1.2 Re-Sequencing Using Non-allele Specific PCR and MiSeq	.38
2.2 Mice	.41
2.2.1 Mouse Strains used in the <i>Sleeping Beauty</i> Study	.41
2.2.2 Transplant of NSG mice	.41
2.2.3 Mice in the <i>PiggyBac</i> Study: Cloning <i>Vk*hPB</i> and <i>Vk*MYC-TA-hPB</i>	.42
2.2.4 Genotyping Transgenic Mice	.44
2.3 Sample Collection and Processing	46
2.3.1 Collection and processing of blood samples from live mice	.46
2.3.2 Necropsy of sick mice, sample collection and processing	.46
2.3.3 Processing of live cells	47
2.3.4 Generation of single cell derived haematopoietic colonies for transplant	.48

A Study of Molecular Synergy and Clonal Evolution in Haematopoietic Malignancies

2.3.5 Preparation of Metaphase Spreads and FISH analysis48
2.3.6 DNA extraction
2.3.7 Exome Sequencing of Mouse SB Tumours
2.3.8 Comparitive Genomic Hybridisation (CGH)49
2.3.9 RNA extraction
2.4 Sequencing transposon integration sites: the Roche 454 Method50
2.4.1 Splinkerette PCR to identify transposon integration sites
2.4.2 Transposon mapping and common integration site (CIS) analysis of 454 data52
2.4.3 Detecting Intra-GrOnc Jumping using PCR, Splinkerette and Sequencing55
2.5 Illumina Sequencing of Transposon Integrations
2.5 Illumina Sequencing of Transposon Integrations582.5.1 Library Preparation58
2.5.1 Library Preparation58
2.5.1 Library Preparation582.5.2 Transposon mapping and CIS analysis of Illumina data60
2.5.1 Library Preparation582.5.2 Transposon mapping and CIS analysis of Illumina data602.6 Additional methods for the Vk*MYC-TA-hPB and Vk*hPB models61
 2.5.1 Library Preparation
 2.5.1 Library Preparation

3. Whole exome sequencing reveals rapid acquisition of driver mutations and branching evolution in a case of NPM1 positive CMML transforming to AML

3.1 Introduction	67
3.2 Clinical Case	68
3.3 Results	70
3.4 Discussion	77

4. Sleeping Beauty driven leukaemogenesis follows a rapid Darwinian-like evolution in a mouse model of Npm1c+ acute myeloid leukaemia

4.1 Introduction	83
4.2 Results	85
4.2.1 <i>Npm1^{cA}</i> mutant mice with a low copy number Sleeping Beauty transposon develop myeloid leukaemias	85
4.2.2 GRL verifies CISs identified by GRH and identifies additional ones	89
4.2.3 <i>Sleeping Beauty</i> driven leukaemia develops suddenly without detectable antecedent abnormalities in the peripheral blood	92

A Study of Molecular Synergy and Clonal Evolution in Haematopoietic Malignancies

4.2.4 Transposon mobilisation begins early and continues throughout the pre-leukaemic pe	eriod 94
4.2.5 A small number of transposon integrations occur early and persist in the pre-leuka samples and on serial transplantation of leukaemia cells	
4.2.6 CIS in the pre-leukaemic blood samples	105
4.2.7 Some transposons loose the capacity to re-mobilise	106
4.2.8 Searching for alternative drivers in transposon IM mice	108
4.3 Discussion	109

5. Development and validation of a protocol for quantitative analysis of transposon integrations

5.1 Introduction
5.2 Results
5.2.1 The TraDIS Illumina Sequencing Protocol Generates High Coverage and Quantitative Data121
5.2.2 TraDIS Identifies Additional CIS Compared to Restriction-Based Mapping122
5.2.3 PCR duplicate removal decreases the proportion of reads attributed to the top hits but does not significantly alter ranking of integration sites
5.2.4 Integrations that persisted on serial sampling generally had high read coverage using TraDIS
5.3 Discussion

6. PiggyBac insertional mutagenesis of the mature B cell compartment

6.1 Introduction	۱	157
6.2 Results		162
6.2.1 Cloning	Vk*HPB and Vk*MYC-TA-HPB	162
6.2.2 Validation	ion of splicing in the transgenic constructs	162
6.2.3 The <i>Vk*</i>	*MYC-TA-HPB construct generates an active PB transposase: HAT resista	nce assay
	B transposase is active <i>in vivo</i> , although transposon mobilisation is not lir I compartment	
6.2.5 Insertion	nal mutagenesis mice have increased lymphoma-associated mortality	166
Vk*-Myc-T	A-hPB mice	168
Vk*-hPB m	nice	169
6.2.6 Immund	ophenotyping to determine developmental stage of the B cell tumours	172
6.2.7 The MY	C-TA-hPB tumours are not universally MYC dependent	178
6.2.8 Stop cod	don reversion was not seen in <i>Vk*hPB</i> and <i>Vk*MYC-TA-hPB</i> tumours	
	B and MYC-TA-hPB IM tumours are clonal and have undergone somatic	
hypermutatio	on	184
6.2.10 Serum	protein electrophoresis of MYC-TA-HPB and HPB mice	

A Study of Molecular Synergy and Clonal Evolution in Haematopoietic Malignancies

6.2.11 Common integration site analysis identifies known and novel lymphoma genes	191
6.2.12 Read depth and correlation with sample clonality	
6.3 Discussion	200

7. Discussion

7.1 Transposon IM as a tool for cancer gene discovery	.210
7.2 Transposon IM as a tool for studying clonal evolution	.215
7.3 Concluding remarks	. 225

8. References	
9. Appendices	