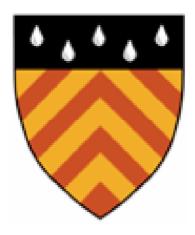
The Transcriptional Profile of Microglia: From Brain to Dish



Fiona Elizabeth Calvert

Clare Hall

December 2019

University of Cambridge

This thesis is submitted for the degree of Doctor of Philosophy

Declaration of originality

This thesis is the result of my own work and includes nothing which is the outcome of

work done in collaboration except as declared in the Preface and specified in the

text. It is not substantially the same as any that I have submitted, or, is being

concurrently submitted for a degree or diploma or other qualification at the University

of Cambridge or any other University or similar institution except as declared in the

Preface and specified in the text.

I further state that no substantial part of my thesis has already been submitted, or, is

being concurrently submitted for any such degree, diploma or other qualification at

the University of Cambridge or any other University or similar institution except as

declared in the Preface and specified in the text.

It does not exceed the prescribed word limit for the relevant Degree Committee

Fiona Calvert

December 2019

3

The Transcriptional Profile of Microglia: from Brain to Dish Fiona Elizabeth Calvert

Microglia are the tissue resident macrophages of the central nervous system (CNS) and multiple lines of evidence indicate that microglia are a pathogenic cell type in Alzheimer's disease (AD). It is important to understand the transcriptional profiles of microglia, both from primary human cells and the *in-vitro* model systems used to study the cells at scale. In this thesis, I aim to build on previous small-scale studies of primary microglia and *in-vitro* model systems to answer three major questions: 1. Can transcriptional data from fresh, primary human microglia be used to identify novel subpopulations of cells and understand how clinical phenotypes influence gene expression? 2. How accurately do current simple *in-vitro* model systems of human microglia capture the profile of primary human cells? 3. Do more complex model systems move cultured cells further along a trajectory towards the primary cell type?

I have utilised RNA-sequencing technology to build the most comprehensive transcriptional profile of primary human microglia to date, from over 100 neurosurgical patients. Using single-cell sequencing I have demonstrated that clinical pathology, particularly major trauma, causes specific gene expression changes within microglial transcriptomes. I have then shown that in-vitro models of primary microglia have significantly reduced expression of key marker genes and transcription factors, such as P2RY12 and SALL1, when compared to primary cells. Using gene-set enrichment analysis tools, I have shown that many of the genes with higher expression in primary cells can be linked to neuronal processes such as CNS myelination. Data from the third chapter of this thesis identified the CNS environment as a major stimulating factor in the gene expression profile of primary microglia. Therefore, I used single cell analysis to understand how culturing stem cell derived microglia in the presence of neurons could move in-vitro systems closer towards the primary cell type. In summary, the work in this thesis has demonstrated that microglial transcriptomes are constantly reacting to stimuli within the local CNS environment, both to maintain their unique gene expression profiles and to respond to clinical conditions. I have also shown that current in-vitro model systems do not fully capture this transcriptional profile which largely appears to be driven by environmental stimuli within the CNS.

Acknowledgments

I sat staring at this page for a little while before writing this, I was unsure of how to put into words the thanks and gratitude I have for all the people who have helped me get to the point of completing this. I also have a tendency to overdo the soppy and felt pressured to write something witty and lighthearted. What follows, much like my PhD, I'm sure will be different from what I expected and planned but something that I will be proud of nonetheless.

First of all, I have to acknowledge Dan Gaffney, my ever wonderful and patient supervisor. I think this PhD has been a learning curve for both of us but I cannot stress enough how vital a part you have played in my journey here. You have pushed and challenged me in ways I couldn't have imagined when I started over four years ago. Throughout it all, you have also provided constant support and encouragement and have never made me feel like I couldn't do this. Thank you for taking a student who didn't even know the terminal existed and only knew how to perform a t-test and shaping me into an R-loving scientist with an annoying obsession for correct statistics. I would also like to acknowledge the wonderful lab you have built, a group of people who have been such a huge part of my PhD. To Andy, who acted as my supervisor within the lab and is truly one of the most remarkable scientists I have ever met. You made me a better scientist and opened my eyes to the wonderful world of moths and orchids. I hope you know that the Gaffney lab is only what it is, in part, because of you. To Julie and Clara, who I could not have survived the last couple of months without, thank you for listening to my gripes and moaning and for always giving the best life advice. To everyone else in the Gaffney lab (Natsuhiko, Nikos, Beata, Maria P, Maria I and Gerda) thank you for providing so many laughs and thoughtful discussions over lunch and coffee breaks. It has been a pleasure to work alongside you all and I hope our GIF-filled slack channels will live on forever!

To everyone I have had the pleasure of working with in these four years outside of the PhD - thank you for providing me with the most wonderful distractions when the PhD took its toll. To the members of staff at Clare Hall, who helped us put on an event that is one of my proudest achievements. The May Ball was a shining light of my four years. To everyone I have met through the Story Collider, the producers and the storytellers, you are the most inspiring people I have ever met. You have changed the way I view science and have opened my eyes to a whole new wonderful world. Particularly to Erin, Liz and Steve - you have taught me so much in such a short space of time and I honestly feel privileged every day that I get to work with such ridiculously amazing people.

I could not have gotten to the point of finishing this PhD without my personal support network. To Lindsay, who I met at the Sanger interviews and knew we would be friends instantly - THANK YOU. You guickly became one of my best friends and I will always be grateful to have someone to cry in the toilets with. I write this full of pride that we both made it through, I wouldn't be here without you. To all my friends outside the PhD, both old and new: Emily, Becky, Emma, Madi, Guy, Luke, Sarah and Christy - thank you for providing wonderful relief from this process and filling my life with joy and excitement. My parents have always been, and I'm sure will continue to be, my biggest cheerleaders. I owe everything I have achieved to the strength and encouragement you have given me. I am so proud to be your daughter and don't know how to ever thank you for all you have done for me. I promise to only make you read this small part of my PhD and will not enforce proof-reading duties on you, even though I know you would if I asked. I love you both so much. Finally, the biggest acknowledgment of all. To Will, my long suffering partner who has been by my side for the majority of this PhD. You have had to listen to me complain about every failed experiment, every bug in my code and every frustration. You have seen me at my worst and provided me with every bit of support I need to pull myself out of those dark spaces. You made me laugh every day and have shown me how to not take life too seriously, something I desperately needed. You have been my rock, my escape and my source of happiness throughout every up and down of this PhD. I cannot thank you enough for being part of my life and for learning what microglia are for me.

Lindsay - we had many discussions about this very point so I include this as an acknowledgement of our journey. A large part of this acknowledgement goes to me, this thesis will forever be a reminder of what I can achieve even when I don't think I can.

Table of contents

Abbreviations	15
Chapter 1: Introduction	17
1.1 Identification and characterisation of microglial cells in the brain	17
1.2 Lineage of microglial populations in the brain	18
1.2.1 Microglial cell origin in embryonic development	18
1.2.2 Maintenance of microglial populations throughout adulthood	19
1.3 Microglial function in development and the adult brain	21
1.3.1 The role of microglia in the developing brain	22
1.3.2 Microglia in adulthood	23
1.4 Microglia and disease	24
1.4.1 Microglia in traumatic brain injury	25
1.4.2 Microglia in Multiple Sclerosis	26
1.4.3 Microglial response in other neurological disorders	27
1.5 Alzheimer's disease and microglia	28
1.5.1 Early hypotheses in Alzheimer's disease research	29
1.5.2 Alzheimer's disease genetics and the neuroinflammation hypothesis	31
1.5.3 The role of microglia in Alzheimer's disease	36
1.6 Studying human microglia	38
1.6.1 Transcriptomic studies in primary human microglia	39
1.6.2 Modelling human microglia	40
1.7 Thesis overview	42
Chapter 2: Heterogeneity in primary adult microglial transcriptomes	45
2.1 Introduction	45
2.1.1 Marker gene identification in mice and human samples	46
2.1.2 Fresh, primary human microglia bulk RNA-sequencing	46
2.1.3 Single cell sequencing and primary microglia	47
2.1.4 The impact on age and sex on microglial transcriptomes	48
2.2 Methods	50
2.2.1 Experimental design and sample collection	50
2.2.2 Tissue processing and cell sorting	51
2.2.3 RNA handling	52
2.2.4 Initial processing and quality control of sequencing data	56
2.2.5 Comparison of bulk data to publicly available datasets	56
2.2.6 Classification of microglial cells using publicly available datasets	57
2.2.7 Variance components analysis	57
2.2.8 Clustering of single cell data, differential expression and cline metadata links	nical 58
2.2.9 Pathway enrichment analysis	58

	2.3 Quality control analysis across datasets	59
	2.3.1 Bulk RNA-sequencing quality control	59
	2.3.2 Metadata comparison	62
	2.4 Single cell clustering and identification of sub-populations	64
	2.4.1 Comparison to publicly available single cell datasets	64
	2.4.2 Clustering of microglial cells and cluster maker analysis	66
	2.5 Clinical metadata and microglial transcriptome signatures	70
	2.5.1 Variance components analysis	70
	2.5.2 Gene expression linked to clinical metadata	71
	2.6 Microglia and disease	76
	2.6.1 Microglial gene expression and Alzheimer's disease (AD)	76
	2.7 Discussion	78
Cł	apter 3: Comparison of in-vitro models of microglia	81
	3.1 Introduction	81
	3.1.1 Monocyte-derived macrophages	82
	3.1.2 Cancer cell lines	82
	3.1.3 iPSC derived macrophages	83
	3.1.4 iPSC derived microglia	84
	3.1.5 Limitations of current transcriptional comparisons across model syste	
	3.2 Methods	85
		86 86
	3.2.1 Data collection and initial processing	88
	3.2.2 Principal components and variance components analysis3.2.3 Differential expression and gene set enrichment analysis	90
	3.3 Technical comparisons within the dataset	91
	3.3.1 Normalisation comparison	91
	3.3.2 Variance components analysis	93
	3.3.3 Effects of differing gene set inputs on principal components analysis	94
	3.4 Utilising principal component analysis to identify sources of variation	96
	3.4.1 Defining principal components	96
	3.4.2 Varimax analysis of principal components	99
	3.5 Differential expression between cell types	101
	3.5.1 Primary microglia vs all models	101
	3.5.2 Primary microglia vs individual model systems	103
	3.5.3 iPSC macrophages vs iPSC microglia	105
	3.6 Expression of Alzheimer's disease genes across model systems	107
	3.6.1 Expression of known Alzheimer's disease genes	107
	3.6.2 Expression of late onset Alzheimer's disease linked genes	110
	3.7 Discussion	113
Cł	apter 4: Complex in-vitro model systems	117
	4.1 Introduction	117

4.1.1 Co-culture and organoid model systems	118
4.1.2 Single cell sequencing and developmental trajectory inference	119
4.2 Methods	120
4.2.1 Cell culture, dissociation and sorting	120
4.2.2 Bulk sequencing preparation	121
4.2.3 Single cell sequencing preparation	122
4.2.4 Bulk RNA-sequencing data processing and analysis	123
4.2.5 Single cell RNA-sequencing data processing and quality control	124
4.2.6 Cluster identification, differential expression analysis and traject analysis	ctory 124
4.3 Bulk RNA-sequencing comparison of complex and simple model systems	125
4.3.1 Dimensionality reduction	125
4.3.2 Differential expression analysis	132
4.4 Identification and clustering of myeloid cells within the single cell dataset	135
4.4.1 Clustering analysis to identify myeloid cells within the full population	135
4.4.2 Partition and cluster analysis using Monocle3	137
4.4.3 Partition marker genes	140
4.5 Cell trajectory analysis across model systems	144
4.5.1 Creation of the trajectory graph	144
4.5.2 Gene expression changes along pseudotime	146
4.6 Discussion	148
Chapter 5: Discussion	151
5.1 Sequencing primary human microglia	151
5.2 Modelling primary microglia in-vitro	153
5.3 Studying microglia in Alzheimer's disease	155
5.4 Concluding remarks	157
References	159

Abbreviations

Transcripts per million

Yolk sac

Uniform Manifold Approximation and Projection

Variance stabilisation transformation

AD Alzheimer's disease Blood brain barrier BBB **CNS** Central nervous system CSF-1R Colony stimulating factor 1 receptor Embryoid body ΕB Expression quantitative trait loci eQTL Fluorescence-activated cell sorting **FACS** Genome-wide association studies **GWAS iPSCs** Induced pluripotent stem cells Knockout KO Late onset Alzheimer's disease LOAD Log fold change **LFC** Monocyte derived macrophages **MDMs** Multiple sclerosis MS Nuclease free water NFW Peripheral Blood Mononuclear Cells **PBMCs** Principal components analysis **PCA** Polymerase chain reaction **PCR** Quality control QC Quantile normalisation QN Single cell RNA-sea scRNA-seq Single nucleotide polymorphism **SNP** Traumatic brain injury TBI TF Transcription factor

TPM

VST

YS

UMAP