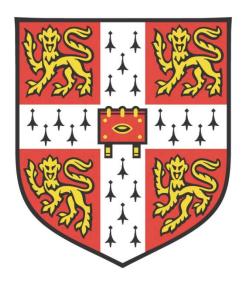
The Genetic Architecture of Immune-Mediated Complex Diseases



Jimmy Zhenli Liu

Darwin College University of Cambridge

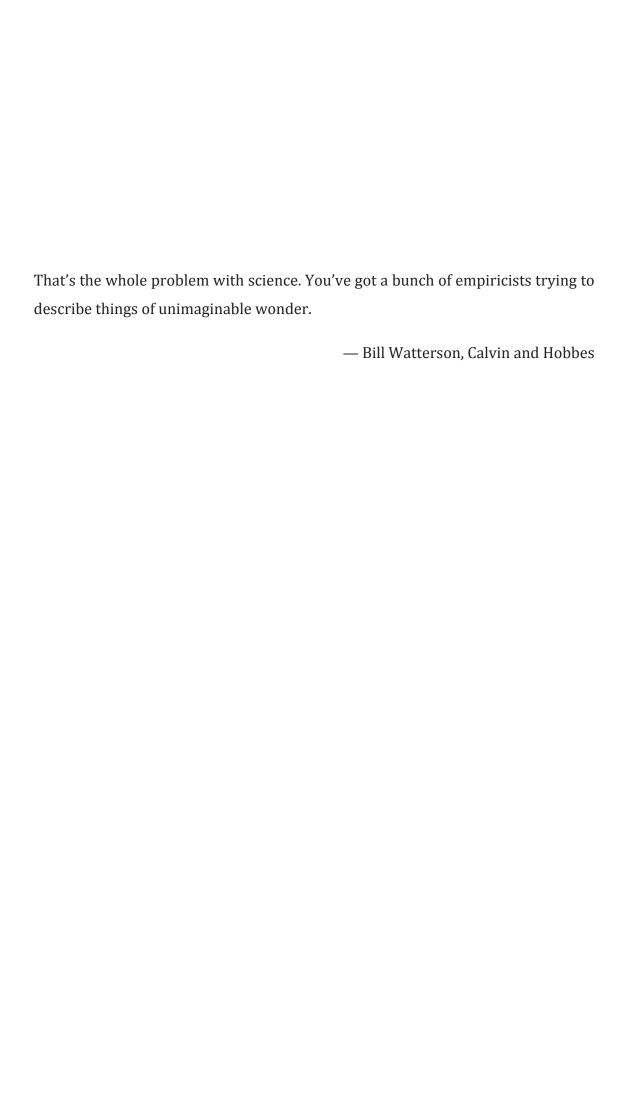
This dissertation is submitted for the degree of Doctor of Philosophy September 2014

Declaration

This dissertation is the result of my own work and includes nothing which is the outcome of work done in collaboration except where specifically indicated in the Contributions section within each chapter. It does not exceed the word limit set by the Degree Committee for the Faculty of Biology, and is not substantially the same as any work that has been, or is being, submitted to any other university degree, diploma or any other qualification.

Jimmy Liu

18 September, 2014



Abstract

Complex disease risk is characterised by a combination of multiple genetic factors along with the environment. Since 2005, genome-wide association studies have discovered thousands of genetic variants associated with hundreds of such diseases. Following on from these types of studies, custom genotyping arrays with dense SNP content have allowed for greater refinement across risk loci, while their low cost has enabled powerful locus discovery projects and cross-phenotype comparisons in very large sample sizes. Combining risk loci with disease-relevant functional genomic data allows for insights into the biology of disease. In this dissertation, I explore locus discovery, cross-phenotype comparisons and functional data integration across four immune-mediated complex diseases – primary biliary cirrhosis, primary sclerosing cholangitis, and the two forms of inflammatory bowel disease – Crohn's disease and ulcerative colitis.

In Chapter 1, I provide a historical background of our understanding of how genetic variation contributes to phenotypic variation, and the technological and theoretical advances in the last twenty years that have lead to the large-scale high-throughput locus discovery projects of today.

In Chapter 2, I describe a locus discovery project using the Immunochip custom genotyping array for primary biliary cirrhosis. In addition to identifying three new risk loci and refining associated variants within known risk loci, I explore how integrating association results with functional genomic annotations across various cell lines from the ENCODE Project can provide insights into the cell types and genomic features most relevant to disease.

In Chapter 3, I describe a similar locus discovery project using the Immunochip for primary sclerosing cholangitis (PSC), where nine novel risk loci were identified. Over 80% of PSC patients are also diagnosed with inflammatory bowel disease, the majority of which is ulcerative colitis. I explore genetic factors

that may explain this overlap, and show that despite this high comorbidity, around half of PSC risk loci appear unique to PSC.

In Chapter 4, I describe a trans-ethnic genome-wide association metaanalysis for inflammatory bowel disease (IBD) comprising individuals of European, East Asian, Indian and Iranian ancestry genotyped on a combination of genome-wide arrays and the Immunochip. Forty new IBD loci were discovered associated with Crohn's disease, ulcerative colitis or both. I show that there exists pervasive sharing of IBD risk loci between European and non-European populations, while also noting specific loci where effect sizes differ between populations. The study demonstrates the utility of performing large-scale GWAS meta-analyses across different populations to identify novel susceptibility loci.

I then move beyond locus discovery in Chapter 5, where I describe a simple method for integrating differential gene expression datasets with disease risk loci. I applied the method to two gene expression datasets reflecting the genes that are involved in maintaining intestinal T cell homeostasis, and those triggered in the gut in response to infection. I find that in both cases, genes that are differentially expressed between these conditions are significantly overrepresented among risk loci for a range of autoimmune disorders, allowing for the identification of additional candidate genes at these loci and the generation of hypotheses about the mechanism through which they mediate disease.

Finally, in Chapter 6, I discuss the major themes of the preceding chapters on unravelling the genetic architecture of complex diseases. I then look to the types locus discovery projects that will shape the field in the coming years, and the potential for these to be ultimately translated into better treatment outcomes for patients.

Acknowledgements

First and foremost, I thank my supervisor, Carl Anderson, for giving me the opportunity to pursue this PhD. It has been an absolute privilege. This dissertation would not have been possible without your continued guidance, enthusiasm and ceaseless faith in me throughout these years. With your stubborn attention to detail and unrelenting loyalty to rigour, you stand as a role model scientist for myself and no doubt many others to come.

I also thank my secondary supervisor, Jeff Barrett, and my degree committee, Stephen Sawcer and Ines Barroso for their guidance over these years. Thank you also to Christina Hedberg-Delouka, Annabel Smith, Alex Bateman and Julian Rayner for keeping the Sanger PhD program such a well-oiled machine.

To members of the Anderson Group (both past and present) - Tejas Shah, Eva Serra, Sun-Gou Ji and Jamie Floyd - I could not have asked for nicer folks to share an office with. It's been an absolute pleasure working with you all; thank you for putting up with me.

The work presented in this dissertation would not have been possible without the efforts of collaborators both at Sanger and around the world, of which there are far too many to list here. But for their hard work, dedication and willingness to share the spoils of research, I am especially indebted to Mohammed Al Marri, Luke Jostins, Daniel Gaffney, Richard Sandford, Trine Folseraas, Tom Hemming Karlsen, Johannes Roksund Hov, Eva Ellinghaus, Andre Franke, Tim Raine, Adam Reid, Suzanne van Sommeren, Rinse Weersma and Hailiang Huang. Thank you also to legions of doctors, nurses, researchers and administrators of the UK PBC Consortium, the International PSC Genetics Consortium and the International IBD Genetics Consortium for their tireless efforts in bringing together groups around the world towards the common noble goal of advancing disease research. None of this of course would have been possible without the >100,000 donors whose DNA were used in these projects, for which I will be forever grateful.

I would also like to thank the many friends and colleagues I've gotten to know during my time at Sanger. Chris Franklin, Yang Luo, James Morris, Scott Shooter, Isabelle Cleynan, Mari Niemi and everyone in Morgan N333 and N309 - cheers for the lunchtime banter, post-lunchtime strolls, post-stroll tea breaks, mingles and thoughts. To everyone I've enjoyed playing with (and competing against) in the Genome Campus volleyball, football and cricket leagues, I tried my best and can only apologise. Thank you to the fellow PhD students and Sanger and those I've gotten to know in Cambridge for making my time here so enjoyable.

I also thank the Wellcome Trust for generously funding my time at Sanger, as well as the various funding bodies for making these projects possible.

Lastly, I want to thank my family, especially my parents, Hua and Xiaoyu, for the unconditional love, support, understanding and encouragement that they have shown me in everything that I do.

Table of Contents

Abstract		iv
Acknowled	gements	vi
Publication	S	xii
From t	his dissertation	xii
Arising	g elsewhere	xii
List of table	es	xiii
List of figur	es	xiv
Chapter 1.	Introduction and historical perspective	1
1.1 In	nmune-mediated diseases	2
1.1.1	The immune system	2
	Epidemiology	
1.2 G	enetic studies of complex autoimmune disorders	4
1.2.1	Mendelian inheritance, multifactorial traits and heritability.	4
1.2.2	Twin studies	8
1.2.3	The major histocompatibility complex	9
1.2.4	Linkage	9
1.2.5	Candidate genes	11
1.2.6	Genome-wide association studies	12
1.3 In	sights from GWAS	16
1.3.1	Biology	16
1.3.2	Genetic overlap between immune-mediated disorders	17
1.4 Lo	ocus discovery beyond GWAS	18
1.4.1	Dense genotyping	18
1.4.2	Finemapping and inferring causality	19
1.4.3	Sequencing and rare variant associations	20
1.5 Co	onclusions	23
1.6 0	utline of dissertation	23
•	Discovery, refinement and functional genomics integrati	
primary	biliary cirrhosis risk loci using the Immunochip	26

2.1 In	troduction	26
2.1.1	Chapter overview	27
2.1.2	Contributions	28
2.2 Me	ethods	28
2.2.1	Samples, DNA extraction and genotyping	28
2.2.2	Quality control	29
2.2.3	Imputation	31
2.2.4	Association analysis	31
2.2.5	HLA Imputation	31
2.2.6	Variance in disease risk explained	32
2.2.7	eQTL analysis	32
2.2.8	Enrichment of open chromatin regions	33
2.3 Re	sults and discussion	34
2.3.1	Replicating known PBC risk loci	34
2.3.2	Multiple independent signals	36
2.3.3	Novel PBC risk loci	39
2.3.4	Associations with HLA haplotypes	40
2.3.5	Functional annotations and enrichment of open chromatin	
	regions among risk loci	41
2.4 Co	nclusion	46
Chapter 3.	Discovery of primary sclerosing cholangitis risk loci and t	he
genetic i	relationship with inflammatory bowel disease	48
3.1 In	troduction	48
3.1.1	Chapter overview	49
3.1.2	Contributions	49
3.2 Me	ethods	49
3.2.1	Samples, DNA extraction and genotyping	49
3.2.2	Quality control	50
3.2.3	Imputation	51
3.2.4	Association analysis	53
3.2.5	Functional annotation of risk loci	54
3.2.6	GRAIL and DAPPLE analyses	55

3.2.7	HLA imputation and association analysis	55
3.2.8	Heritability explained	55
3.2.9	Prediction of PSC using IBD risk loci	56
3.2.10	Genetic correlation between PSC and IBD	56
3.3 Re	esults and discussion	57
3.3.1	Locus discovery	57
3.3.2	Associations at previously reported non-HLA PSC risk loci	61
3.3.3	Candidate gene prioritisation	62
3.3.4	HLA association	63
3.3.5	Genetic overlap with IBD	66
3.4 Co	onclusion	73
Chapter 4.	Trans-ethnic meta-analysis for inflammatory bowel disea	ıse
risk loci	and population comparisons	75
4.1 In	troduction	75
4.1.1	Contributions	76
4.2 M	ethods	77
4.2.1	Sample collection and genotyping	77
4.2.2	Immunochip quality control	77
4.2.3	Per-population association analysis	80
4.2.4	Transethnic meta-analysis	80
4.2.5	Gene prioritisation	82
4.2.6	Variance explained	82
4.2.7	Heterogeneity of effect sizes and allele frequencies between	
	populations	82
4.2.8	Genetic correlation	83
4.2.9	Gene-based likelihood ratio test	83
4.3 Re	esults and discussion	85
4.3.1	Per-population association and transethnic meta-analysis	85
4.3.2	Candidate genes	87
4.3.3	Validation of known loci	90
4.3.4	Population comparisons	92
4.3.5	Gene-based likelihood ratio test	98

4.3.6	Conclusions	100
Chapter 5.	Immune-mediated disease risk loci are enriched for	
differe	ntially expressed genes from tissue-relevant functional	
genom	ic datasets	102
5.1 In	ntroduction	102
5.1.1	Contributions	105
5.2 M	lethods	105
5.2.1	Human T cell transcripts	105
5.2.2	Mouse cecum transcripts	106
5.2.3	GWAS enrichment	106
5.3 R	esults	107
5.3.1	Human T cell transcripts	107
5.3.2	Mouse cecum transcripts	109
5.4 D	iscussion	111
5.4.1	Conclusions	115
Chapter 6.	Conclusions and future prospects	117
6.1 E	ffect sizes, power and the genetic architecture of comple	ex
t	raits	118
6.2 F	uture prospects for complex disease genetics	123
6.2.1	Array-based approaches	123
6.2.2	Sequencing approaches for rare variant studies	124
6.2.3	Genetic studies in non-European populations	129
6.2.4	Genetic prediction	130
6.3 F	rom causal variants to treatment outcomes	131
6.4 C	oncluding remarks	135
Bibliograp	hy 136	

Publications

From this dissertation

- Liu J.Z., van Sommeran S., Huang H., Ng S.C. *et al.*, Association study discovers 38 susceptibility loci for inflammatory bowel disease and shows pervasive sharing of genetic risk across diverse populations. *Under review*.
- Raine T., Liu J.Z., Anderson C.A., Parkes M. and Kaser A., Generation of primary human intestinal T cell transcriptomes reveals differential expression at genetic risk loci for immune-mediated disease. Gut. *In press.*
- Liu J.Z. and Anderson C.A., Genetic studies of Crohn's disease: past, present and future. Best Practice & Research: Clinical Gastroenterology, 28:373-386, 2014.
- Foth B.J., Isheng J.T., Reid A.J., Bancroft A., *et al.*, Whipworm genomes and dual-species transcriptome analysis provide molecular insights into an intimate host-parasite interaction. Nature Genetics, 46:693-700, 2014.
- Liu J.Z., Hov J.R., Folseraas T., Ellinghaus E., *et al.*, Dense genotyping of immune-related disease regions identifies nine new risk loci for primary sclerosing cholangitis. Nature Genetics 45:670-675, 2013.
- Liu, J.Z., Almarri, M.A., Gaffney, D.J., Mells, G.F., Jostins, L., Cordell, H.J., Ducker, S.J., Day, D.B., Heneghan, M.A., Neuberger, J.M., *et al.* Dense fine-mapping study identifies new susceptibility loci for primary biliary cirrhosis. Nature Genetics, 44:1137-1141, 2012.

Arising elsewhere

- Curtis J., Luo Y., Zenner H.L., Cuche-Lourenco D., *et al.*, Susceptibility to tuberculosis is associated with the ASAP1 gene that regulates dendritic cell migration. *Under Review*.
- Houldcroft C.J., Petrova V., Liu J.Z., Frampton D., *et al.*, Host genetic variants and gene expression patterns associated with Epstein-Barr virus copy number in lymphoblastoid cell lines. PLoS One 9:e108384, 2014.
- Robles-Espinoza C.D., Harland M., Ramsay A.J., Aoude L.G., *et al.*, POT1 loss-of-function variants predispose to familial melanoma. Nature Genetics, 46:478-481, 2014.
- Shah T.S., Liu J.Z., Floyd J.A.B., Morris J.A., *et al.*, optiCall: A robust genotyping-calling algorithm for rare, low frequency and common variants. Bioinformatics 28:1598-1603, 2012.

List of tables

Table 2.1. Sample quality control30
Table 2.2. SNP quality control30
Table 2.3. Unconditioned and conditioned association results for the four independent signals at 3q25
Table 2.4. Genome-wide significant HLA-type associations
Table 3.1. Post-QC patient and control panels50
Table 3.2. Association results of twelve non-HLA genome-wide significant risk loci for PSC
Table 3.3. Association of genome-wide significant PSC risk loci with other diseases
Table 3.4. Candidate functional annotations and genes among genome-wide significant PSC risk loci
Table 3.5. Stepwise conditional analyses of the classical HLA genes64
Table 3.6. IBD Subphenotypes among PSC cases
Table 4.1. Post-QC patient and control panels genotyped on the Immunochip 78
Table 4.2. Post-QC case and control panels used in the transethnic meta-analysis
Table 4.4. Candidate genes implicated by coding variants, eQTLs, GRAIL and DAPPLE in 28 of the 40 novel IBD risk loci
Table 4.5. New genes in known IBD risk loci implicated from GRAIL and DAPPLE network analyses89
Table 4.6. Pairwise genetic correlation (r _G) tagged by Immunochip SNPs94
Table 4.7. Genes that exceeded $P < 5 \times 10^{-5}$ in at least one non-European cohort in the likelihood ratio locus-based test
Table 5.1. Enrichment of genes that are upregulated in gut T cells compared with blood T cells in loci associated with six phenotypes108
Table 5.2. Enrichment of genes that are differentially expressed between infected and uninfected cecum tissue among loci associated with six phenotypes109
Table 5.3. Annotation of disease-associated loci that are show nominal levels of enrichment ($P < 0.05$) for genes that show differential expression in healthy gut vs. blood T cells and in infected vs. uninfected mouse cecum tissue110

List of figures

Figure 1.1. Mendel's laws of inheritance5
Figure 1.2. Polygenic inheritance in a normally distributed trait: height6
Figure 1.3. The liability threshold model
Figure 1.4. Power of linkage vs. association outlined in Risch and Merikengas (1996)14
Figure 1.5. Number of publications indexed in PubMed with the terms "autophagy" and "Crohn's" in the abstract since 2006
Figure 2.1. Principal component analysis of PBC cases and controls30
Figure 2.3. PBC risk loci odds ratios from this study vs. those from Mells <i>et al.</i> (2011)
Figure 2.4. Multiple independent signals at 3q25 from stepwise conditional regression
Figure 2.5. Enrichment of DNase-seq peaks among PBC risk loci in Gm12878 compared to other ENCODE cell lines
Figure 2.6. Enrichment of DNase-seq peaks among PBC risk loci calculated from P-value bins
Figure 3.1. Heterozygosity rate and proportion of missing genotypes for PSC cases and controls
Figure 3.2. Principal components analysis of PSC cases and controls with 1000 Genomes Omni2.5-8 data
Figure 3.3. Quantile-quantile plots and genomic inflation factors of observed vs. expected P-values
Figure 3.4. Regional association plots for genome-wide significant associations at previously established PSC risk loci
Figure 3.5. Regional association plots of nine newly associated PSC risk loci 60
Figure 3.6. Regional association plots from stepwise conditional regression in the HLA complex in PSC
Figure 3.7. Odds ratio comparisons for PSC risk loci in IBD. IBD ORs and designation of loci as UC, CD or both (IBD) were obtained from Jostins <i>et al.</i> (2012)
Figure 3.8. Venn diagram of directions of effect in PSC of SNPs associated with either CD, UC or both (IBD)
Figure 3.9. Predicting PSC using OR estimates from CD and UC risk loci68
Figure 3.10. Predicting the IBD subphenotypes of PSC patients using OR estimates from CD and UC risk loci

Figure 3.11. Genetic correlation (r_G) estimates using genome-wide SNP data between CD/UC and PSC subphenotypes70
Figure 3.12. Two models of pleiotropy
Figure 3.13. Odds ratios of PSC risk loci calculated using all PSC cases compared with odds ratios calculated using PSC+UC and PSC+no IBD subphenotypes 73
Figure 4.1. Principal components analysis of non-European IBD patients and controls
Figure 4.2. Comparison of samples used in this study with those from Jostins et al. (2012)80
Figure 4.3. GRAIL network for all genes with GRAIL P < 0.05
Figure 4.4. Comparison of association P-values reported in Jostins <i>et al.</i> (2012) and Europeans in this present study
Figure 4.5. Odds ratio comparison between European and non-European populations at 233 SNPs associated with CD, UC other both
Figure 4.6. Belgravia plot of (A) CD and (B) UC risk variants in Europeans and East Asians96
Figure 5.1. Number of upregulated genes that overlap among CD4+ LPL, CD8+ LPL, CD4+ IEL and CD8+ IEL T cells vs. counterparts in blood
Figure 5.2. Quantile-quantile plots of gene length of differentially expressed genes in (A) gut T cells vs. blood and (B) infected vs. uninfected cecum tissue.114
Figure 6.1. Effective sample size vs. number of genome-wide significant risk loci across GWAS and Immunochip studies of nine immune-mediated disorders119
Figure 6.2. Cumulative proportion of variance in disease liability explained by the genome-wide significant loci identified in Chapters 2-4120
Figure 6.3. The genetic architecture of inflammatory bowel disease126

