Computational Analysis of Genomes

Matthew R. Pocock

This dissertation is submitted for the degree of Doctor of Philosophy.

April 2003

Supervisors: Dr T. J. P. Hubbard, Dr N. Goldman

The Sanger Centre, Cambridge; Darwin College, Cambridge

This dissertation is the result of my own work and includes nothing which is the outcome of work done in collaboration.

The work in this thesis has not been submitted in whole, or in part, for a degree, diploma, or any other qualification at any other university.

Matthew R. Pocock,

April 2003, Cambridge, United Kingdom

Dedication

I would like to thank all of those who have supported me through the process of

producing this dissertation. Particular thanks must go to Tim Hubbard, who has been

a source of great help and provided direction where needed without smothering me

with micro-management. Nick Goldman and Ed Griffiths have both been valuable

sounding boards throughout. Thomas Down has been my staunchest ally as we

developed BioJava and also the DAS protocol from embryonic beginnings to the well-

respected projects they are now. It would be unfair not to thank all of those who have

helped with BioJava, as coders, testers and users. In particular, a mention must go to

Chris Dagdigian for managing the hardware. I have enjoyed my time here, and this is

in no small part due to the friendliness of those I meet daily in the Sanger Centre, the

EBI and from the Ensembl project. Lastly, I must dedicate this work with heartfelt

gratitude to Caroline. Without her love all academic achievements would be

worthless.

Fate is unmoved by one's pitiful hopes; what changes, bowing to fate, is

what one hopes for.

(Liza Dalby, The Tale of Murasaki p239)

ii

Abstract

Recently we have been blessed with a simultaneous rise in the volume of biological data and the power of computers. This has necessarily led to the emergence of the field of Bioinformatics, where the study of entire genomes rather than individual genes is the norm.

This dissertation describes the development and application of the software framework BioJava, designed from the outset to provide a strong foundation for the implementation of different machine learning algorithms. BioJava allows genomic size datasets to be efficiently manipulated in a range of hardware environments.

A variety of supervised and unsupervised learning techniques were applied to data sets on the scale of whole genomes taking advantage of the BioJava framework.

Firstly, unsupervised learning was used to look for underlying structure in the genome sequence of whole Malaria chromosomes. Time-reversible 1st order Hidden Markov Models (HMMs) learned signals based on sequence composition that appear to correlate closely with biological units, such as exons, introns, repeats and non-coding genomic regions. This demonstrates the ability of unsupervised methods to discover biologically meaningful information within genomic sequence.

Secondly, supervised learning was used to develop a regression method able to predict recombination rate within human chromosomes. Support Vector Machines (SVMs) using suffix tree kernels were trained on human chromosome 22 sequence and were able to learn a signal reproducibly, although it was not clear how well this models recombination rate.

Finally, supervised learning was used to develop a classification method able to detect subtle signals in noisy and small sets of micro-array expression data. A Bayesian technique for training linear models was applied to learn sparse models. These were able to distinguish between tumour samples that had been treated with a drug and those that had not. The models produced by this method can be readily interpreted in terms of individual genes, and in this case made good biological sense.

This dissertation illustrates how a framework of modular and reusable software components can be used together with advances in artificial intelligence to help us interpret the data flowing from high throughput projects in the post genomic era.

Table of Contents

Dedication	11
Abstract	iii
Table of Contents	v
Table of Figures	ix
List of Tables	xi
Table of Equations	xii
Chapter 1 Introduction	1
1.1 Existing Software Development Frameworks for Bioinformatics	2
1.1.1 The NCBI Toolkit	3
1.1.2 Bioperl	4
1.1.3 EMBOSS	5
1.2 BioJava	6
1.3 Machine Learning	8
1.3.1 Clustering, Classification and Regression for Single Items	9
1.3.2 Signal Analysis with Hidden Markov Models	18
1.4 Implementation and Use of BioJava	24
Chapter 2 The BioJava Core Interfaces	24
2.1 Java as a Language for Bioinformatics	24
2.2 Nested Exceptions and Assertions	24

	2.3	Changeability	24
	2.4	Symbols, Alphabets and SymbolList	24
	2.5	Locations, Sequences and Features	24
	2.6	Probability Distributions and Hidden Markov Models	24
	2.7	Query	24
	2.7.	1 Motivations	24
	2.7.	2 Initial Implementation	24
	2.7.	3 Limitations of This System	24
	2.8	Recent Developments	24
	2.8.	1 The Tag-Value Parser	24
	2.8.	2 Flat File Indexing	24
	2.8.	3 Annotation Types	24
	2.8.	4 Enhanced Feature Filters	24
	2.8.	5 Change Hubs	24
	2.8.	6 Bit Packed Sequences	24
	2.9	Conclusions	24
C	Chapter	3 HMMs for whole <i>Plasmodium Falciparum</i> Chromosomes	24
	3.1	Introduction	24
	3.2	Simple HMM Architectures	24
	3.2.	1 Methods	24
	3.2.	2 Results	24

3.3 HMN	A Architectures with Complementary Emission Distributions	24
3.3.1	Methods	24
3.3.2	Results	24
3.4 First	Order HMMs with Time-Reversible Transition Probabilities	24
3.4.1	Methods	24
3.4.2	Results	24
3.5 Discu	ussion	24
3.6 Futur	re Directions	24
Chapter 4 Inv	vestigation of Recombination Rates Using SVMs	24
4.1 Intro	duction	24
4.1.1	Support Vector Machines	24
4.1.2	BioJava APIs for Support Vector Machines	24
4.2 Meth	ods	24
4.2.1	Searching for a Signal Affecting Recombination Rates Using a	
Word-Fre	quency Kernel Function	24
4.2.2	Construction and Training of an SVM for Predicting	
Recombin	ation Rate	24
4.3 Resu	lts	24
4.3.1	Recombination Rates Predictions	24
4.3.2	Cross-Validation	24
4.4 Disci	ussion	2.4

Chapter 5	RVMs for Classification of Expression Data	24
5.1	Introduction	24
5.2	Cellular Responses to Doxorubicin	24
5.3	Generalized Linear Models	24
5.4	Micro-array Classification Using a Support Vector Machine Imple	mented
as a Lin	near Kernel RVM	24
5.4.1	Framework for Generalised-Linear-Models amenable to	
Expr	ession Arrays	24
5.4.2	RVM Analysis Using the Small Working Set Heuristic	24
5.4.3	Function of Genes Identified by GLM Models	24
5.5	Conclusions, Applications and Future Work	24
Concludin	ng Remarks	24
Reference		24

Table of Figures

Figure 3-1 Emission probabilities for the four pair-state model	24
Figure 3-2 Diagram of the <i>P. Falciparum</i> chromosome 3 and the state paths through	;h
three models	24
Figure 3-3 Diagram of the <i>P. Falciparum</i> chromosome 2 and the state paths through	;h
three models	24
Figure 3-4 Emission Spectrums for all Pair-State Models	24
Figure 3-5 Diagram of the alignments of the 3,4 and 5 state-pair models to Malaria	
chromosome 3	24
Figure 3-6 Diagram of the alignments of the 3,4 and 5 state-pair models to Malaria	
chromosome 2	24
Figure 3-7 Counts for Biological Feature and States for the 2-5 Pair-State Models	24
Figure 3-8 Normalized Counts of States for Biological Features	24
Figure 3-9 Normalized counts of Biological Features for States	24
Figure 4-1 Comparison of physical and genetic distances along chromosome 22	24
Figure 4-2 Total Results of Training the SVM using Uniform Counts	24
Figure 4-3 Moving Average for Uniform Counts models of Depth 4-6	24
Figure 4-4 Moving Average for Uniform Counts models of Depth 7-9	24
Figure 4-5 Total Results of Training the SVM using Normalized Rates	24
Figure 4-6 Moving Average for Normalized Rates: Depths 4-6	.24
Figure 4-7 Moving Average for Normalized Rates: Depths 7-9	24

Figure 4-8 Accuracy for Recombination SVMs Under 3-Way Jack-knifing	24
Figure 4-9 Predictions Across the Entire Chromosome from the 3 Jack-knife Mod	dels
for Depth of 5	24
Figure 5-1 Scatter Plot of the Two Topoisomerase II Probes Used.	24
Figure 5-2 Expression Levels for Each Probe Used	24
Figure 5-3 Average Weighs Across Relevant Models	24
Figure 5-4 Average Weights Across All Models	24

List of Tables

Table 3-1 Forward-strand and reverse-strand counts	24
Table 3-2 State-transitions and their reverse-complements	24
Table 5-1 GLM for all before-after pairs (to 4 s.f.)	24
Table 5-2 Genes used by cross-validation models	24

Table of Equations

Equation 1-1 A Hypothesis Function.	10
Equation 1-2 Error of a Hypothesis	11
Equation 1-3 Some Error Functions	12
Equation 1-4 Dot Products for Items Decomposable into Sub-Spaces with Dot-	
products Defined	14
Equation 1-5 Definition of Kernel Functions	15
Equation 1-6 A Polynomial From a Two-dimensional Coordinate to a Coordinate	
Containing One Component for each Possible Product Involving up to Two	
Dimensions	15
Equation 1-7 Dot products between two polynomial mappings reduced to terms	
involving the dot product of the unmapped variables	16
Equation 1-8 Polynomial Kernel Function	16
Equation 1-9 Definition of a Probabilistic Hidden Markov Model	21
Equation 1-10 Emission and Transition Probabilities	22
Equation 1-11 Definition of All Legal State-Sequences	23
Equation 1-12 Likelihood of Observing a Given Sequence and Labelling	23
Equation 1-13 Common Dynamic Programming Recursions as Applied to	
Probabilistic Hidden Markov Models	24
Equation 4-1 Equation of a Plane	24
Equation 4-2 Normal to a Plane as a Weighted Sum of Vectors	24

Equation 4-3 Definition of a Support Vector Machine	24
Equation 4-4 Basis Functions for Kernel Functions and Data Points	24
Equation 4-5 SVMs in Terms of Basis Functions	24
Equation 4-6 The Normalizing Kernel	24
Equation 4-7 SuffixTree Kernel	24
Equation 5-1 Bayes Theorem	24
Equation 5-2 Rearrangement of Bayes Theorem	24
Equation 5-3 Bayes Theorem in Words	24