Identification of Extrachromosomal Elements from Whole Genome Sequences of the Human Gut Microbiome to Investigate the Gut Mobilome and Resistome





Tapoka Thulisile Mkandawire USN:303635303

University of Cambridge

Murray Edwards College

This dissertation is submitted for the degree Master of Philosophy in Biological Science @

The Sanger Institute

Table of Contents

1. Introduction 1.1 The Gut Microbiome 5 1.2 Residents of the Gut Microbiota 5 1.3 The Role of the Microbiota in Health and Disease 7 1.4 Plasmid Biology 8 1.5 Plasmid Replication and Maintenance 10 1.6 Horizontal Gene Transfer 11 1.7 Frequency of and Challenges to HGT 13 1.8 The Utility of Plasmids as Tools for Biotechnology 14 1.9 Plasmids and the Gut Microbiome 15 1.10 Plasmid Functions in the Gut Microbiome 16 1.11 HGT in the Gut Microbiome 17 1.12 Plasmid Capture Strategies 18 1.13 Challenges to Plasmid Capture 19 1.14 Hypothesis 20 1.15 Project Aims 20 1.16 Project Rationale and Identified Challenges 20 2. Methods 2.1 Computational Plasmid Isolation, Phenotypic Predictions, and Phylogeny 22 2.2 Bacterial Culture and Growth Curve Plotting 23 2.3 Growth Medium 24 2.4 Species Validation by 16S rRNA gene PCR 24 2.5 Plasmid Extractions and Digests 25 2.6 Plasmid Visualisation by Gel Electrophoresis 25 2.7 Antimicrobial Sensitivity Testing 25 2.8 Transformation 25 2.9 Conjugation 26 3. Bioinformatics Results 3.1 Frequency of Plasmid Detection Across Human Gut Microbiota Phyla 27 3.2 Size and Coverage and Distribution of Predicted Plasmids 29 3.3 Plasmid Classification and Phylogeny 30 3.4 AMR Gene Distribution 35 4. Experimental Results 4.1 Plasmid Isolation 37 4.2 Species Validation 37 4.3 Plasmid Digests 38 39 4.4 Antimicrobial Screening 4.5 Strain Growth Monitoring 40 4.6 Plasmid Transfer 41 5. Discussion 5.1 Distribution of Plasmids in the Human Gut Microbiome Culture Collection 45 5.2 Genomic Inference of Biological Functions: Plasmid Classification and AMR Distribution 46 5.3 Transformation of Culture Collection Isolates 48 5.4 Applications of Identified Plasmid Sequences 49 5.5 Plasmids and Plasmid Sequence Data as Genetic Tools 49 5.6 Strategies to Engineer the Microbiota 51 5.7 Challenges to Bacteriotherapy 53 5.8 Non-Health Associated Uses of Engineered Microbiota 54 5.9 Strategies for Investigating Engineered Microbiota 54 5.10 Summary

List of Figures and Tables

1. Introduction 1.1. Figure: Distribution of bacterial phyla and environmental composition along the GIT 6 1.2. Figure: Phylogenetic tree of the Lawley lab human gut microbiota culture collection 6 9 1.3. Figure: Diagram of the conformations of plasmids 10 1.4. Figure: Summary of the process of rolling circle replication 1.5. Figure: Figure summarising the methods of horizontal gene transfer 11 1.6. Figure: Phylogenetic tree displaying the distribution of bacteria in environmental water 13 1.7. Figure: Chart displaying the frequency with which a variety of vectors are used in gene therapy trials 15 1.8. Figure: Graph displaying the distribution of RelE genes in the gut microbiome 16 1.9. Figure: Flow diagram outlining the variety of processing routes of stool samples for 19 plasmid analysis 2. Methods 2.1. Figure: Workflow diagram summarising the strategy taken to probe the plasmids of the human gut microbiome 22 3. Bioinformatic Results 3.1. Figure: The distribution of predicted plasmids in the 653 genomes of the Lawley Laboratory culture collection 28 3.2. Figure: Lawley Lab culture collection phylogeny with plasmid prediction annotations 29 3.3. Figure: Detected plasmid coverage compared to chromosomal median coverage and plasmid distribution by size 30 3.4. Figure: Phylogenetic tree constructed using the *repE* gene 31 3.5. Figure: Phylogenetic tree constructed using the repA gene 32 3.6. Figure: Chord diagram displaying links between genomes containing 33 closely related repE genes. 3.7. Figure: Chord diagram displaying links between genomes containing closely related repA genes 34 3.8. Figure: Percentage bar graph displaying the proportions of predicted resistance 36 4. Experimental Results 37 4.1. Figure: Agarose gel displaying the isolated plasmids greater than 10kb 4.1. Table: Results of the 16S species validation 38 4.2. Figure: Agarose gel displaying small plasmids in native (U) and digested (D) form 39 4.2. Table: Antibiotic resistance levels as observed with Etest strips 39 4.3. Figure: Agarose gel displaying mid size and large plasmids in native (U) and digested (D) form. 39 4.4. Figure: OD growth curves for selected isolates 41 4.5. Figure: Results of the conjugation between the donors (D) and naladixic acid engineered recipient (R) 42 4.6. Figure: Results of the conjugation between the donors (D) and erm-resistant bacteria (R) 43 4.7. Figure: Results of the conjugation between donor H5 29 and aerobic bacterial recipients 44 5. Discussion 5.1. Figure: The three key synthetic circuits: dynamic, logic, and communication circuits 52

Acknowledgements

I'd like to thank my supervisor Dr Trevor Lawley and the members of Team 162: Anne, Sam, Nitin, Lindsay, Yan *et al.*, for all the support and input. As well as my thesis committee for their invaluable insights: Dr David Summers, Prof Nick Thomson.

Abstract

We are covered with beneficial microbial communities, termed microbiota, that play important roles in our health, sustenance and well-being. Pathological imbalances in our microbiota caused by things like antibiotic exposure and poor diet can directly cause, or predispose us to, a variety of diseases and metabolic syndromes. Our intestinal microbiota contains hundreds of uncultured bacterial species and the genetic complement of our microbiota, our microbiome, contains 150 times more genes than the human genome. The human microbiome is dynamic due to bacteria's ability to horizontally transfer large blocks of DNA between distantly related species. Horizontal gene transfer by mobile elements such as transposons and plasmids plays a central role in the evolution and functions of wellstudied intestinal bacteria such as Escherichia coli and Lactobacillus. However, most of the human microbiota has never been cultured and characterized so the composition of mobile elements in the majority of the human microbiota remains poorly defined. For my MPhil Thesis, I analysed a large scale human commensal reference genome collection of 653 genomes, representing the phylogenetic diversity of the human intestinal microbiota, for the presence of extrachromosomal elements like plasmids and transposons. A combination of bioinformatics analyses and wet-lab validation methods have been employed to identify, isolate, and characterize elements from gut bacteria creating a catalogue of known and novel sequences. 240 genomes were predicted to contain extrachromosomal DNA, these elements are primarily small and high coverage, and predicted to contain resistance genes. A small number of megaplasmids were also detected, and a phylogeny was built to identify any plasmids with a broad host range. This database can be used as a reference for the computational isolation of extrachromosomal elements from whole genome sequencing data and metagenomic datasets. They will also form the foundation for developing tools for genetic manipulation of the novel and uncharacterized gut commensal microbes of the culture collection.