Genome-wide recessive screens for DNA mismatch repair genes in mouse ES cells
This dissertation is submitted for the degree of Doctor of Philosophy.
By Zikai Xiong
The Wellcome Trust Sanger Institute Hughes Hall, University of Cambridge

DECLARATION

I hereby declare that this dissertation is the result of my own work and includes nothing which is the outcome of work done in collaboration, except where specifically indicated in the text.

None of the material presented herein has been submitted previously for the purpose of obtaining another degree.

Zikai Xiong

ACKNOWLEDGEMENT

I would like to take this opportunity to formally thank all the people who supported me during the course of my PhD project.

My deepest thank is to my PhD supervisor Dr Allan Bradley, for his enormous support and excellent advice through the whole project. It was great pleasure to work with Allan and achieve challenging tasks in his laboratory. I thank his patience as well as encouragement during the hard time of this project.

I have special thanks to all Bradley laboratory members for their communication and kind help. I am particularly grateful to my wife Dr Wei Wang for her huge support in both my life and career. We have had numerous scientific discussions and collaborations during these experiments. I learned recombinant retroviral techniques from her. Many thanks are also due to Oliver Dovey for his detailed guidance and contribution to array CGH experiments. I also want to thank Peter Ellis for his help on expression arrays and I am grateful to Gregory Lefebvre, Keith James and Robert Andrews for their help in bioinformatics analysis. I thank Dr Ge Guo and Dr Wei Wang (Male) for their instructions in ES cells' culture and molecular manipulations and valuable discussions. I also thank Dr Yue Huang for discussions at the beginning of the project. I thank Haydn Prosser for providing vectors and advice, Qi Liang for comfirming the recombination medicated cassette exchange ability of my cell line in her experiments. I am grateful to Frances Law and Alastair Beasley for their help in cells' culture and routine laboratory issues. I thank Shaun Cowley, Juan Cadinanos, Roland Rad, George Vassiliou, Antony Rodriguez, Nathalie Conte for valuable scientific discussions.

The advice and feedback from my PhD committee were precious. The members were Pentao Liu, Jessica Downs, Andrew Fraser, Derek Stemple and Allan Bradley, who shared their critical thinking and provided excellent advice for my project.

I must say "Thank you!" to my parents Yan Xiong and Huiqin Zhu. Their endless support never leaves me.

My 4-year full studentship is sponsored by the Wellcome Trust. Registered charity number 210183

ABSTRACT

Genome-wide genetic screens on libraries containing homozygous mutant mouse embryonic stem (ES) cells make it feasible to examine all mouse genes expressed in ES cells for their role in any biological process which is active in ES cells. Bloom syndrome protein (Blm) deficient ES cells have high mitotic recombination and loss of heterozygosity rates, which allow cells with homozygous mutations to be generated in populations of mutated *Blm*-deficient ES cells. A genome-wide mutation library was generated in this genetic background using gamma radiation. Analysis of isolated clones from this library by high-resolution Comparative Genomic Hybridization (CGH) arrays revealed that each carried several duplications or deletions ranging in size from 0.1 to 50Mb. This mutation library provided good coverage of the mouse genome and it is a new genetic resource for conducting loss-of-function genetic screens in mammalian cells.

In humans, mutations in components of the DNA mismatch repair (MMR) system cause hereditary non-polyposis colorectal carcinoma (HNPCC, Lynch syndrome). Mutations in the *MSH2* and *MLH1* genes account for the majority of the cases. To extend our understanding of the MMR system a recessive screen was implemented in the mutation library to identify new components by selection in 6-thioguanine (6TG). This nucleotide analogue is incorporated into DNA and is recognized by the DNA MMR complex, leading to cell death in wild type cells while MMR-deficient cells are viable. In this screen, several independent mutants with homozygous deletions covering the MMR genes *Msh2* and *Msh6*, and one mutant with a heterozygous deletion at the *Dnmt1* gene were isolated. Mutants were also isolated in which a number of unknown genes were deleted. Joint analysis using genomic and transcriptional arrays discovered that a set of genes, including *Msh2* and *Msh6*, were deleted and silenced in the 6TG^R mutants. The most frequently silenced genes are highly likely to be involved in the MMR pathway.

This screening system confirmed that irradiation is efficient in generating genome-wide loss-of-function mutation libraries in the *Blm*-deficient cells, and combined with DNA- and RNA-based array analysis platforms, provides a new approach for phenotype-driven recessive genetic screens in mouse ES cells.

TABLE OF CONTENTS

LIST OF FIGURES)
LIST OF TABLES14	1
ABBREVIATIONS	;
CHAPTER 1. INTRODUCTION	,
1.1 The mouse as a genetic model animal	17
1.1.1 Evolution divergence of the mouse and the human	17
1.1.2 The history of the laboratory mouse	17
1.1.3 Mouse genomics	18
1.2 Mouse embryonic stem cells as genetic research tools	21
1.3 Genetic screens	22
1.4 Strategies to make homozygous mutations in ES cells	24
1.4.1 Sequential gene targeting	24
1.4.2 High concentration of G418 selection	26
1.4.3 Induced mitotic recombination	29
1.4.4 Blm gene deficiency elevates the rate of loss of heterozygosity	33
1.4.4.1 Bloom syndrome	33
1.4.4.2 Cloning of the <i>BLM</i> gene in humans	33
1.4.4.3 DNA helicase activity of BLM	34
1.4.4.4 Proteins interacting with BLM	34
1.4.4.5 Mouse models of Bloom syndrome	36
1.4.4.6 Increased rate of loss of heterozygosity in Blm-deficient mouse ES cell	
applications	
1.5 Experimental mutagens used in mouse genetics	
1.5.1 N-ethyl-N-nitrosourea	46
1.5.2 Ionizing radiation	
1.5.2.1 Effects on DNA	
1.5.2.2 Applications of ionizing radiation in mouse genetics	
1.5.3 Gene targeting	
1.5.4 Insertional mutagenesis systems	
1.5.4.1 Gene-trap mutagenesis	
1.5.4.2 Retroviruses	
1.5.4.3 Transposon-mediated mutagenesis	
1.6 DNA mismatch repair	
1.6.1 DNA mismatch repair in prokaryotes	
1.6.2 DNA mismatch repair in eukaryotes	
1.6.3 MMR in hereditary non-polyposis colon cancer	
1.6.4 Mouse models for human DNA mismatch repair gene defects	
1.6.5 MMR deficiency and tolerance of DNA methylation	/ /

1.6.6 6-thioguanine introduces DNA mismatches, cycle arrest and apoptosis	s 78
1.6.7 Genetic screen for MMR genes	82
1.6.8 Project design	82
CHAPTER 2. MATERIALS AND METHODS	85
2.1 Cell culture	85
2.1.1 Mouse ES cell culture	85
2.1.2 Blm-deficient ES cell line	85
2.1.3 Dnmt1-deficient ES cell line	87
2.1.4 Chemicals and media used for ES cell culture	87
2.1.5 Passaging ES cells	90
2.1.6 Freezing ES cells	90
2.1.7 Electroporation of DNA into ES cells	91
2.1.8 Picking ES cell colonies	91
2.1.9 Production of recombinant retrovirus	91
2.1.10 Recombination Mediated Cassette Exchange	92
2.2 Gamma Irradiation	94
2.3 DNA methods	94
2.3.1 Solutions used in molecular experiments	94
2.3.2 PCR	95
2.3.3 Primers	95
2.3.4 Transformation of DNA into E. coli strains	96
2.3.5 Recombineering	97
2.3.5.1 Principle	97
2.3.5.2 Recombineering protocol	100
2.3.6 Vectors	103
2.3.6.1 pZK5 – <i>Puro∆tk/Kanamycin</i> cassette	103
2.3.6.2 pZK9 – Vector to retrieve gene desert DNA from BAC	103
2.3.6.3 pZK15 – Retrieving the homology required for targeting from a B	AC 106
2.3.6.4 pZK8 - Chromosome 6 gene desert mini-targeting vector	106
2.3.6.5 Construction of gene desert targeting vector pZK-GD	109
2.3.6.6 RMCE plasmid with the <i>Bsd</i> cassette	112
2.3.6.7 Retrieve part of <i>Blm</i> gene for targeting from a BAC	112
2.3.6.8 Retrieval product with <i>Blm</i> gene exons 2 and 3 (pZK19)	112
2.3.6.9 Blm gene mini targeting vector (pZK31)	115
2.3.6.10 Blm gene-targeting vector (pZK30)	117
2.3.7 Genomic DNA isolation from tissue culture (6-well plate)	120
2.3.8 Genomic DNA isolation from 96-well plates	120
2.3.9 Endonuclease digestion for Southern blotting	120
2.3.10 Southern blot hybridization	
2.3.10.1 Probes	
2.3.10.2 Southern blotting	121
2.3.10.3 Probe preparation	121

2.3.10.4 Hybridization and membrane washes	122
2.3.11 Array comparative genomic hybridization	122
2.3.11.1 Principle	122
2.3.11.2 Reagents	124
2.3.11.3 Random labelling DNA for array CGH	125
2.3.11.4 Hybridization	125
2.3.11.5 Array washing	126
2.4 RNA methods	126
2.4.1 RNA isolation	126
2.4.2 Expression arrays - RNA isolation and sample labelling	127
2.5 Statistical analysis of arrays	129
2.5.1 Array CGH	129
2.5.2 Expression array	129
2.5.2.1 Detection P-value	129
2.5.2.2 Combined P-value and transcripts' presence	
2.5.2.3 Comparison analysis	130
CHAPTER 3. EVALUATING SIZES OF HOMOZYGOUS DELETIONS 131	
3.1 Introduction	131
3.1.1 Gene deserts	131
3.1.2 A strategy to isolate mutants with homozygous deletions	135
3.2 Results	
3.2.1 Chromosome 6 gene desert knock-in cell ZK2.1	137
3.2.2 Testing background FIAU resistance levels prior to selection for homozygous deletions.	139
3.2.3 Puromycin and FIAU double resistant clones contain the <i>Puro∆tk</i> cassette	
3.3 Discussion	146
CHAPTER 4. ATTEMPT TO GENERATE A ${\it Blm}$ -DEFICIENT CELL LINE WITI	H BAC
ACCEPTOR 148	
4.1 Introduction	148
4.2 Results	150
4.2.1 Generation of the ZK6 wild type BAC acceptor cell line	150
4.2.2 Confirmation that RMCE can be achieved	154
4.2.3 Blm ^{+/-} BAC acceptor cell lines	154
4.2.4 Using Flpe/FRT to remove the selection marker	158
4.2.5 Generation of the BIm ^{neoF} allele	162
4.3 Discussion	164
CHAPTER 5. CELL VIABILITY AND CHROMOSOME REARRANGEMENTS A	AFTER
GAMMA IRRADIATION166	
5.1 Introduction	166
5.2 Results	
5.2.1 Cell viability after irradiation	

5.2.2 Irradiation generates megabase sized chromosomal alterations	174
5.3 Discussion	187
5.3.1 Survival frequency of ES cells after irradiation	187
5.3.2 Megabase duplications and deletions generated by irradiation	187
5.3.3 Chromosomal abnormalities	188
CHAPTER 6. PILOT MISMATCH REPAIR SCREEN	
6.1 Introduction	189
6.2 Results	191
6.2.1 The procedure of the pilot screen	
6.2.2 The complexity of mutant pools	193
6.2.3 Genomic changes after irradiation and FIAU selection	193
6.2.4 Efficiency of gamma irradiation	195
6.2.5 Clone ITC8 was 6TG-resistant	195
6.2.6 ITC8 carries a homozygous deletion covering the Msh2 and Msh6 loci	197
6.3 Discussion	199
CHAPTER 7. GENOME-WIDE MISMATCH REPAIR SCREEN201	
7.1 Introduction	201
7.2 Results	204
7.2.1 Use of MMuLV to provide molecular tags in mutation pools	204
7.2.2 Gamma-irradiation mutation library	204
7.2.3 Assessment of screen background	206
7.2.4 6TG-resistant clones	208
7.2.5 Clonal relationship of 6TG ^R clones	217
7.2.6 Array CGH analysis	226
7.2.6.1 Msh2 and Msh6 homozygous deletions	226
7.2.6.2 Msh2 and Msh6 heterozygous deletions	239
7.2.6.3 <i>Msh6</i> mutant G10	
7.2.6.4 A common deletion on chromosome 14	251
7.2.6.5 The Other mutant clones	
7.3 Conclusion and discussion	
7.3.1 Advantages of establishing clonal relationships	
7.3.2 Mutations were identified in <i>Msh2</i> and <i>Msh6</i> MMR genes	
7.3.3 Clues of potential MMR genes from a common deletion	268
CHAPTER 8. Expression analysis of the 6TG ^R mutants	
8.1 Introduction	269
8.2 Results	269
8.2.1 Expression analysis	269
8.2.1.1 Controls of wild type cell lines – AB1 and AB2.2	270
8.2.1.2 Comparison between <i>Blm</i> -deficient and wild type cells	275
8.2.1.3 Comparison between <i>Dnmt1</i> -deficient and AB2.2 wild type cells	284
8 2 1 4 Msh2 and Msh6 homozygous mutants	287

8.2.1.5 Msh6 homozygous deletion in mutant D1	293
8.2.1.6 Expression status of <i>Msh2</i> and <i>Msh6</i> in heterozygous deletions	297
8.2.2 Combined analysis of array CGH and expression array results	301
8.2.2.1 Positive relationship between copy number and expression variation	301
8.2.2.2 Genes affected by MMR defects	304
8.2.2.3 <i>Dnmt1</i> mutant B7	314
8.3 Conclusion and discussion	320
8.3.1 Confirmation of Msh2, Msh6 and Dnmt1 deficiency in mutants	320
8.3.2 Genes regulated by Msh2 and Msh6	321
8.3.3 Potential genes involved in the MMR pathway	321
CHAPTER 9. GENERAL DISCUSSION	
9.1 Blm-deficient ES cells for genetic screens	323
9.2 Selecting for deletions with HSV-tk	325
9.3 Gamma irradiation as an efficient mutagen	326
9.4 Mutation detection methods	328
9.5 Future analysis of the MMR mutants	330
9.6 Other mutagenesis systems	331
REFERENCE 333	

LIST OF FIGURES

Figure 1-1	Conservation of synteny between human chromosome 14 and mouse chromosome 20	: 12 (part)
Figure 1-2	Conserved synteny between humans and mice	20
Figure 1-3	Sequential gene targeting	25
Figure 1-4	High concentration G418 selection facilitates recovery of homozygous mutants	27
Figure 1-5	Induced mitotic recombination	31
Figure 1-6	Mouse Blm gene transcript	37
Figure 1-7	The alignment of the BLM protein sequence in humans and mice – 1/3	37
Figure 1-8	The alignment of the BLM protein sequence in humans and mice – 2/3	38
Figure 1-9	The alignment of the BLM protein sequence in humans and mice – 3/3	39
Figure 1-10	Mutant Blm alleles and mouse phenotypes	41
Figure 1-11	Blm deficiency affects rates of sister chromatid exchange in ES cells	44
Figure 1-12	Mitotic recombination	45
Figure 1-13	Basic gene-trap vectors	56
Figure 1-14	Recombinant retroviral vectors and viral production	60
Figure 1-15	E. coli mismatch repair system	67
Figure 1-16	Human DNA mismatch recognition preferences	70
Figure 1-17	The molecules of guanine and its analogue 6-thioguanine (6TG)	78
Figure 1-18	Cytotoxicity of MNNG and 6TG	79
Figure 1-19	A genome-wide recessive screen for DNA mismatch repair genes	84
Figure 2-1	Genotype of the <i>Hprt</i> ^{+/+} <i>Blm</i> -deficient ES cell line NGG5-3	86
Figure 2-2	Structure of the <i>Dnmt1</i> locus in the targeted cell lines	88
Figure 2-3	The principle of Recombination Mediated Cassette Exchange	93
Figure 2-4	The basic functions of Bacteriophage lambda proteins: Exo and Beta	98
Figure 2-5	The annealing of the ssDNA to the replication fork by lambda Beta protein	99
Figure 2-6	The structure and function of the defective lambda prophage	101
Figure 2-7	Gap repair process	102
Figure 2-8	Vector pZK5 – <i>Puro∆tk/Kanamycin</i> cassette	104
Figure 2-9	Structure of pBS <i>DT-A</i> plasmid	104
Figure 2-10	Vector pZK9 – to retrieve gene desert genomic DNA from a BAC	105
Figure 2-11	Generation of retrieval product pZK15	107
Figure 2-12	Structure of pZK8 – gene desert mini targeting vector	108
Figure 2-13	Generation of the chromosome 6 gene desert targeting vector - pZK-GD	110
Figure 2-14	Restriction pattern confirmed correct pZK-GD plasmids	111
Figure 2-15	Construction of the RMCE vector pZK33	113
Figure 2-16	Blm gene retrieval vector pZK21	113
Figure 2-17	Retrieving Blm gene-targeting arms from a BAC	114
Figure 2-18	Enzyme digestion to identify the correct recombinants	114

Figure 2-19	Generation of the Blm gene mini targeting vector - pZK31	116
Figure 2-20	Generation of Blm targeting vector pZK30	118
Figure 2-21	Screens for the pZK30 by enzyme digestion	119
Figure 2-22	The principle of array comparative genome hybridization	123
Figure 2-23	RNA amplification and labelling for expression array	128
Figure 3-1	A gene desert on chromosome 6	133
Figure 3-2	Conserved linkage between mouse chromosome 6 gene desert region and huma	an genome
	133	
Figure 3-3	Syntenic genes around the chromosome 6 gene desert	134
Figure 3-4	Design to generate homozygous deletions in gene desert region	136
Figure 3-5	Chromosome 6 gene desert targeting	
Figure 3-6	Gene desert targeted clone with resistance to puromycin and FIAU	140
Figure 3-7	Puromycin and FIAU double resistant background in NGG5-3 cells	142
Figure 3-8	Southern blot analysis confirms the presence of <i>Puro∆tk</i> cassette	144
Figure 4-1	Generation of ZK6 BAC acceptor cell line	151
Figure 4-2	Genotype of all ZK6 BAC acceptor clones	153
Figure 4-3	The principle of RMCE in ZK6 cell line	155
Figure 4-4	Blm gene-targeting strategy	156
Figure 4-5	PCR genotype of ZK7A9 ZK8H11 subclones	159
Figure 4-6	FIAU-resistant background of ZK7 and ZK8 cells	160
Figure 4-7	Selection confirmed functional markers in <i>Blm</i> targeting vectors	161
Figure 4-8	PCR screen of ZK15 and ZK16 clones	163
Figure 5-1	Gamma irradiation caused viability change in ES cells	173
Figure 5-2	Array CGH analysis on WT2G2 – a wild type clone surviving 2Gy irradiation	176
Figure 5-3	Array CGH analysis on WT5G4 – a wild type clone surviving 5Gy irradiation	177
Figure 5-4	Array CGH analysis on WT7G2 – a wild type clone surviving 7Gy irradiation	178
Figure 5-5	Array CGH analysis on Blm2G2 – a <i>Blm</i> ^{-/-} clone surviving 2 Gy irradiation	179
Figure 5-6	Array CGH analysis on Blm5G1 – a <i>Blm</i> ^{-/-} clone surviving 5 Gy irradiation	180
Figure 5-7	Array CGH analysis on Blm7G1 – a <i>Blm</i> ^{-/-} clone surviving 7 Gy irradiation	181
Figure 5-8	Array CGH analysis on Blm10G1 – a <i>Blm</i> ^{-/-} clone surviving 10 Gy irradiation	182
Figure 6-1	Pilot screen on <i>Blm</i> ^{-/-} cells carrying selected mutations generated by gamma 190	ı irradiation
Figure 6-2	Phenotype confirmation of ITC clones	196
Figure 6-3	Array CGH analysis of 6TG ^R clone ITC8	198
Figure 7-1	The procedure of a genome-wide MMR screen on the MMuLV-infected, irradia	ted ES cell
library	203	
Figure 7-2	Clean background of the 6TG screen on Blm ^{-/-} cells	207
Figure 7-3	Phenotype of mutant clones	210
Figure 7-4	Phenotype of mutant clones	211
Figure 7-5	Phenotype of mutant clones	212
Figure 7-6	Phenotype of mutant clones	213
Figure 7-7	Phenotype of mutant clones	214

Figure 7-8	Phenotype of mutant clones	215
Figure 7-9	Phenotype of mutant clones	216
Figure 7-10	Design of the Puro∆tk probe and the Southern blot strategy to establish	h clona
relations	ships	218
Figure 7-11	Southern blot to establish clonal relationships of 6TGR clones in mutation pools I	3, C and
D	219	
Figure 7-12	Southern blot to establish clonal relationships of 6TG ^R clones in mutation pools	E and F
	221	
Figure 7-13	Southern blot to establish clonal relationships of 6TG ^R clones in mutation pools 223	G and H
Figure 7-14	Msh2 and Msh6 homozygous mutations	229
Figure 7-15	Array CGH profile of mutant D1 with a homozygous deletion at <i>Msh6</i>	
Figure 7-16	Array CGH analysis of mutant D4	
Figure 7-17	Tile path BACs at <i>Msh2</i> and <i>Msh6</i> loci	
Figure 7-18	Zoomed-in regions of <i>Msh2</i> and <i>Msh6</i> homozygous deletions	
Figure 7-19	Schematic view of <i>Msh2</i> and <i>Msh6</i> homozygous deletions in six mutants	
Figure 7-20	Msh2 and Msh6 heterozygous mutants	
Figure 7-21	Msh2 and Msh6 heterozygous mutants – chromosome 17	
Figure 7-22	Array CGH profile of mutant D6	
Figure 7-23	Tile path BACs at <i>Msh6</i> locus	
Figure 7-24	Array CGH profile of mutant G10	
Figure 7-25	A common heterozygous deletion on chromosome 14	
Figure 7-26	Array CGH profile of mutant B3	
Figure 7-27	Array CGH profile of mutant B7	
Figure 7-28	Array CGH profile of mutant E3	
Figure 7-29	Array CGH profile of mutant E9	
Figure 7-30	Array CGH profile of mutant F1	
Figure 7-31	Array CGH profile of mutant F6	
Figure 7-32	Array CGH profile of mutant G6	
Figure 7-33	Array CGH profile of mutant H1	
Figure 7-34	Array CGH profile of mutant H3	
Figure 7-35	Array CGH profile of mutant H5	265
Figure 8-1	Alignment of probe sequences of MPSV	272
Figure 8-2	Mouse Hprt1 gene expression variation between AB1 and AB2.2 cells	274
Figure 8-3	Expression array probes in the <i>Blm</i> gene	276
Figure 8-4	Expression fold change of the <i>Msh2</i> gene in homozygously deleted mutants	289
Figure 8-5	Expression fold change of the <i>Msh6</i> gene in homozygously deleted mutants	290
Figure 8-6	Cox7a1 is up-regulated in all five Msh2 and Msh6 homozygously deleted mutants co	ompared
with the	Blm-deficient NGG5-3 cell line	292
Figure 8-7	Analysis of the mutant D1, a clone with an <i>Msh6</i> homozygous deletion	294
Figure 8-8	Positive relationship between copy number and expression level	302
Figure 8-9	Genes on chromosome 15 and 17 which was deleted in one mutant and down-red	ulated in

others	313	
Figure 8-10	Array CGH profile of mutant B7	315
Figure 8-11	Array CGH analysis of clone B7 at a zoomed-in region of the <i>Dnmt1</i> locus	317

LIST OF TABLES

Table 1-1	A comparison of mutagens to germ line mutations	47
Table 1-2	DNA mismatch repair (MMR) homologues	69
Table 1-3	Mouse models of deficient mismatch repair genes	76
Table 2-1	Electronic sequence analysis of pZK19 and pZK30	118
Table 4-1	The efficiency of RMCE	153
Table 5-1	ES cell plating efficiency	169
Table 5-2	Wild type ES cell survival rates following 0-10 Gray gamma irradiation	170
Table 5-3	Blm-deficient ES cell survival rates following 0-10 Gray gamma irradiation	171
Table 5-4	Summary of ES cell survival rates following gamma irradiation	172
Table 5-5	Summary of chromosome changes in wild type cells caused by irradiation	183
Table 5-6	Summary of chromosome changes in <i>Blm</i> ^{-/-} cells caused by irradiation	184
Table 5-7	Summary of deletions and duplications in wild type cells and Blm-deficient cells gene	erated
by 2-10	Gray gamma irradiation	186
Table 6-1	6TG ^R clones isolated from pools BTY1 and BTY2	192
Table 6-2	FIAU ^R cells in ATY3 and BTY3	194
Table 6-3	Summary of chromosome changes of random picked clones from pool ATY3 and BTY3	. 194
Table 6-4	The ratio of FIAU-resistant and (Puro and FIAU) double resistant colonies before and	l after
irradiat	ion	196
Table 7-1	Complexity of each mutation pool	205
Table 7-2	6TG resistant mutant clones in the mutation library	209
Table 7-3	Redundant mutant clones and unique mutant clones	225
Table 7-4	Summary of homozygous deletions covering Msh2 and/or Msh6	228
Table 7-5	Summary of heterozygous deletions covering Msh2 and Msh6	245
Table 7-6	Genes in the common heterozygous deletion on chromosome 14	253
Table 7-7	Summary of the mutants analysed using array CGH	255
Table 8-1	Main transcriptional variants between the AB1 and AB2.2 cell lines	273
Table 8-2	Genes expressed in wild type cells twice as much as in Blm-deficient cells	277
Table 8-3	Genes expressed in Blm-deficient cells twice as much as in wild type cells	281
Table 8-4	Genes expressed more than two fold higher in the Dnmt1-deficient cells compared wi	th the
AB2.2	wild type cells	285
Table 8-5	Genes expressed more than two fold higher in the AB2.2 wild type cells compared wi	th the
Dnmt1-	-deficient cells	286
Table 8-6	Down-regulated genes in all five Msh2 and Msh6 homozygously deleted mutants com	pared
with the	e Blm-deficient NGG5-3 cell line	291
Table 8-7	Genes down-regulated in both D1 (Msh6"- deletion) and the five Msh2 and Msh6 det	ficient
mutant	s compared to <i>Blm</i> -deficient ES cells	296
Table 8-8	Expression signal decrease of Msh2 and Msh6 probes in homozygously and heterozyg	jously
deleted	I mutants	298

Table 8-9	Genes down-regulated in ten Msh2/Msh6 deficient mutants compared to Blm-deficient E	S
cells	300	
Table 8-10	Genes which are deleted in one mutant and whose transcripts are down-regulated	in
mutants	compared with the <i>Blm</i> -deficient cells	7
Table 8-11	Genes which are deleted in one mutant and whose transcripts are down-regulated	in
mutants	compared with wild type cells (AB1 and AB2.2)	9
Table 8-12 G	Senes which are deleted in one mutant and whose transcripts are down-regulated in mutan	ts
compare	ed with wild type cells and the <i>Blm</i> -deficient cells31	1
Table 8-13	Genes regulated in both mutant clone B7 and <i>Dnmt1</i> knockout cells	9

ABBREVIATIONS

6TG 6-Thioguanine

BAC Bacterial Artificial Chromosome

Blm Bloom syndrome homolog (human) gene

BME β-Mercaptoethanol

bp base pairs

Bsd Blasticidin resistance gene

Bcyd Blasticidin S HCI

CGH Comparative Genomic Hybridization

DMSO Dimethyl Sulfoxide

Dnmt1 DNA methyltransferase (cytosine-5) 1 gene

DSB Double-Strand Break
ENU N-ethyl-N-nitrosourea
FBS Foetal Bovine Serum

FIAU 1-(2'-deoxy-2'-fluoro-β-D-arabinofuranosyl)-5-iodouracil

GPS Glutamine-Penicillin-Streptomycin

Hprt Hypoxanthine guanine phosphoribosyl transferase 1
HSVtk Herpes Simplex Virus type 1 thymidine kinase

IR **Ionizing Radiation** LET Linear-Energy-Transfer LOH Loss of Heterozygosity Mlh1 mutL homolog 1 (E. coli) Mlh3 mutL homolog 3 (E coli) MMS Methyl Methane Sulfonate MMuLV Moloney Murine Leukemia Virus MNNG N-methyl-N'-nitro-N-nitrosoguanidine

Msh2mutS homolog 2 (E. coli)Msh3mutS homolog 3 (E. coli)Msh4mutS homolog 4 (E. coli)Msh5mutS homolog 5 (E. coli)Msh6mutS homolog 6 (E. coli)

NCBI National Center for Biotechnology Information

NMD nonsense-mediated decay

OMIM Online Mendelian Inheritance in Man

PBS Phosphate Buffered Saline
PCNA Proliferating Cell Nuclear Antigen
PCR Polymerase Chain Reaction
PGK phophoglycerate kinase

Pms1 postmeiotic segregation increased 1 (*S. cerevisiae*)
Pms2 postmeiotic segregation increased 2 (*S. cerevisiae*)

PTC premature termination codon

Puro puromycin N-acetyltransferase gene

RFLP Restriction Fragment Length Polymorphism
RMCE Recombination Mediated Cassette Exchange

SDS Sodium dodecyl sulphate

SIGTR Sanger Institute Gene Trap Resource

SSB Single-Strand Break

SSLP Simple Sequence Length Polymorphism